Inhibition of Advanced Glycation End Products Formation by

*Mangifera indica* Leaf Extract

Kimihisa Itoh\(^1\), Kazuya Murata\(^2\), Nao Sakaguchi\(^3\), Kohei Akai\(^3\), Tomoka Yamaji\(^3\), Kohsuke Shimizu\(^1\), Kaoru Isaki\(^3\), Tetsuya Matsukawa\(^1, 3\), Shin’ichiro Kajiyama\(^3\), Masahiko Fumuro\(^1\), Morio Iijima\(^1, 4\) & Hideaki Matsuda\(^1, 2\)

\(^1\)The Experimental Farm, Kindai University, Wakayama, Japan
\(^2\)Faculty of Pharmacy, Kindai University, Osaka, Japan
\(^3\)Faculty of Biology-Oriented Science and Technology, Kindai University, Wakayama, Japan
\(^4\)Faculty of Agriculture, Kindai University, Nara, Japan

Correspondence: Kimihisa Itoh, The Experimental Farm, Kindai University, 2355-2 Yuasa, Yuasa-cho, Arida-gun, Wakayama 643-0004 Japan. Tel: 81-737-622-953. E-mail: itoh_k@nara.kindai.ac.jp

Received: June 16, 2017 Accepted: July 24, 2017 Online Published: August 12, 2017
doi:10.5539/jps.v6n2p102 URL: https://doi.org/10.5539/jps.v6n2p102

Abstract

The purpose of this study was to examine an inhibitory effect of mango leaf extracts on advanced glycation end products (AGEs) formation and to identify these active ingredients, and also to investigate a relationship between leaves maturation and the inhibitory activity. A methanolic extract of old dark green mango leaf extract (OML-ext) exhibited an inhibitory activity of AGEs formation in nonenzymatic glycation of albumin. The inhibitory activity of OML-ext was attributable to 3-C-β-D-glucosyl-2,4,4’,6-tetrahydroxybenzophenone (1), mangiferin (2) and chlorophyll. Inhibitory effect of young dark reddish brown mango leaf extract (YDL-ext) on AGEs formation was similar to that of OML-ext. The inhibitory activity of YDL-ext was attributable to 1 and 2, in addition, a part of the the activity of YDL-ext due to anthocyanins whose content is highest in young dark reddish brown mango leaves. Considering the amounts of leaves obtained from pruning, old dark green leaves may be a reasonable natural resource for the preparation of ingredients with inhibitory activity of AGEs formation.

Keywords: advanced glycation end products (AGEs) formation inhibition, anthocyanin, chlorophyll, 3-C-β-D-glucosyl-2,4,4’,6-tetrahydroxybenzophenone, glycation, *Mangifera indica*, mangiferin

1. Introduction

Glycation is a non-enzymatic browning reaction caused by amino-carbonyl reactions between reducing sugars and amino groups of proteins and lipids (Tsui-Naito, Saeki, & Hamano, 2009). By glycation of these compounds, advanced glycation end products (AGEs) are irreversibly synthesized in the body (Huebschmann, Regensteiner, Vlassara, & Reusch, 2006), and the accumulation of AGEs in organs is induced by hyperglycemia and is one of the causes of diabetic complications (Sourris, Harcourt, & Forbes, 2009). Moreover, AGEs accumulate in the skin of non-diabetics and are correlated with skin aging (Dyer et al., 1993). The AGEs accumulation in the skin induce cross-linking of collagen and reduce skin degradability and dermal regeneration (Wondrak, Roberts, Jacobson, & Jacobson, 2002). In addition, AGEs induce fibroblast apoptosis by adducting to AGE receptors on the cell (Pageon, Bakala, Monnier, & Asselineau, 2007). These phenomena are also thought to be related to skin aging (Lohwasser, Neureiter, Weigle, Kirchner, & Schuppman, 2006). Therefore, in recent years, the role of AGEs has been increasingly discussed in the skin aging, and the inhibition of AGEs formation can be one of the effective strategies for direct alleviation of the development of novel antiaging cosmeceutical ingredients.

An AGEs formation inhibitor, namely aminoguanidine (Pimagedine®) (Dyer et al., 1993) was under development in U.S.A. as a drug for the treatment of diabetic complications such as diabetic nephropathy, however the clinical trial on aminoguanidine has been discontinued due to adverse reactions such as anemia, liver injury and vitamin B6 deficiency. In order to find new and safe AGEs formation inhibitors from natural resources, pharmacological screening of plant is considered as one of strategies. Hitherto, several plant, such as *Thymus vulgaris* whole grass (Morimitsu, Yoshida, Esaki, & Hirota, 1995), *Chrysanthemum morifolium* and
Chrysanthemum indicum corolla (Tsuji-Naito et al., 2009), Alpinia zerumbet rhizomes (Chompoo, Upadhyay, Kishimoto, Makise, & Tawata, 2011), Derris indica stem bark (Anusiri, Choodej, Chumriang, Adisakwattana, & Pudhom, 2014) and Ribes nigrum fruit (Chen et al., 2014, Xu et al., 2016), have been reported to have AGEs formation inhibitory activity.

In our preceding paper (Itoh et al., 2016), we reported that mango (Mangifera indica Linne) leaf extracts exhibited pancreatic lipase inhibitory activities, and a part of the activity of leaf extract was attributable to C-glucosyl-polyphenols, such as 3-C-β-D-glucosyl-2,4,4′,6-tetrahydroxybenzophenone (1) and mangiferin (2), and that dark green mango leaf which was obtained by summer pruning may be a reasonable natural resource for the preparation of ingredients with lipase inhibitory activity. For finding another utility value of pruned mango leaves, we examined the inhibitory effects of mango leaf extracts on AGEs formation. In this paper, we report AGEs formation inhibitory activity of mango leaf extracts, and also discuss a relationship between leaves maturation and the inhibitory activity.

2. Materials and Methods

2.1 Plant Materials

Three kinds of Mango leaves (old dark green leaf, young dark reddish brown leaf, and young yellow leaf, Figure 1) of M. indica (cv. Irwin) were collected according to the preceding paper (Itoh et al., 2016). In order to describe accurately the color of young mango leaves, we describe the color of the leaves as dark reddish brown in this paper instead of dark brown in the preceding paper (Itoh et al., 2016).

![Figure 1. Photographs of typical mango leaves at various stages of development](image)

2.2 Extraction

Methanolic extracts of young dark reddish brown leaves (this extract is abbreviated as YDL-ext throughout this paper), young yellow leaves (YYL-ext), and old dark green leaves (OML-ext) were obtained according to the preceding paper (Itoh et al., 2016).

2.3 Reagents

Aminoguanidine hydrochloride (Lot #: MKCB3580), Bovine serum albumin (BSA, fraction V, Lot #: SLBQ4710V) and authentic 1 and 2 were purchased from Sigma-Aldrich Japan (Tokyo, Japan). Authentic chlorophyll was purchased from Tokyo Chemical Industry Co., Ltd. (Tokyo, Japan). Other chemical and biochemical reagents were of reagent grade and were purchased from Wako Pure Chemical Industries, Ltd. (Osaka, Japan) and/or Nacalai Tesque, Inc. (Kyoto, Japan) unless otherwise noted.

2.4 In vitro AGEs Formation Inhibition Assay by Incubation of Glucose and Albumin

AGE formation activity was measured according to the method of Shimoda et al. (2011) with minor modification. The test sample was dissolved with dimethyl sulfoxide (DMSO) and diluted with sodium phosphate buffer (PBS, 0.2 M KH2PO4, 0.2 M NaOH, pH 7.2) to a final DMSO concentration of 5% v/v. The reaction mixture of glucose (10% w/v) and bovine serum albumin (BSA, 1% w/v) dissolved in PBS (900 µl) was incubated for 48 h at 60°C in microtube (2 ml) with or without a test solution. After incubation, the reaction mixture was diluted with distilled water (1:7 v/v), the fluorescence (F) associated with AGEs was monitored at an excitation wavelength of 370 nm and emission of 450 nm using a multi-label counter (PerkinElmer 2030 ARVO X4, PerkinElmer Life and Analytical Sciences). Aminoguanidine hydrochloride was used as a reference agent. The inhibitory ratio of the sample was calculated using the following formula:

\[
\text{% inhibition} = 1 - \left( \frac{F\ \text{sample} - F\ \text{sample blank}}{F\ \text{control} - F\ \text{normal}} \right) \times 100
\]
where \( F \) control is the fluorescence with PBS containing glucose and BSA; \( F \) normal is the fluorescence with PBS containing glucose and BSA without incubation (stored at 4°C); \( F \) sample is the fluorescence with sample solution in PBS containing glucose and BSA, \( F \) sample is blank the fluorescence with sample solution in PBS.

Each assay was performed in triplicate (P value < 0.01). IC\(_{50}\) value represents the concentration required to inhibit 50% of AGEs formation by incubation of glucose and albumin.

2.5 Isolation of 3-C-β-D-glucosyl-2,4,4’,6-tetrahydroxybenzophenone (1) and Mangiferin (2)

The OML-ext (5.4 g) was submitted to a silica gel column chromatography with a stepwise gradient elution with CHCl\(_3\)/MeOH (10:0 v/v) to (0:10 v/v). Inhibition of AGEs formation at 25 µg/ml of each fraction was assayed. Fraction I (fr. 1-6) eluted firstly with CHCl\(_3\) showed a significant activity. Fraction I was dark green, and it seemed to contain green pigments, such as chlorophyll as one of active constituents. Fraction II (fr. 7-25) eluted with CHCl\(_3\)/MeOH 30:1 v/v, 10:1 v/v, 5:1 v/v and 0:10 v/v also showed potent activity. Further purification of fraction II by a preparative HPLC led to isolation of 1 and 2 as described in the preceding paper (Itoh et al., 2016).

2.6 Content of Chlorophyll, Total Anthocyanin, 1 and 2 in Mango Leaf Extracts

In this paper, content determination of each compound in leaf extracts was performed in triplicate, and values represent the mean ± standard deviation. Spectrophotometric determination of the content of chlorophyll (mg/g extract) was carried out with 80% acetone aqueous solution containing 2.5 mM sodium phosphate buffer (pH 7.8) according to the method of Porra et al. (Porra, Thompson, & Kriedemann, 1989). The content of total anthocyanin (µg/g extract) in each leaf extract was determined by the following HPLC analysis and shown as µg/g extract in which weight (µg) was calculated as that of an external standard, cyanidin-3-O-glucoside chloride. The extract was dissolved in 10% MeOH aqueous solution containing 10% acetic acid, followed by centrifugation at 12,000 G for 10 min. Resulting supernatant was applied to a HPLC system. The HPLC system consisted of LC-20A pump and SPD-20A photodiode array detector (Shimadzu, Kyoto, Japan). The samples were analyzed by using an Inertsil ODS-3 reverse phase column (4.6 × 150 mm, GL Sciences, Tokyo, Japan) and gradient elution with MeOH aqueous solution at a constant flow rate of 0.8 ml/min. The elution was carried out using linear gradient condition as follows; initial condition was set at 5% MeOH and maintained for 5 min, followed by a linear gradient from 5% to 40% MeOH for 35 min. The column temperature was set at 40°C, and eluted compounds were detected at 520 nm. Total anthocyanins content was determined using total 520 nm peak areas from linear calibration curves made from an external standard, cyanidin-3-O-glucoside chloride. Linear calculation curves in the range of 0.02 to 0.1 nmol were made from the peak areas analyzed at 520 nm, and the correlation coefficient was 0.992. The HPLC determination of 1 and 2 in each leaf extract was described in the preceding paper (Itoh et al., 2016).

2.7 Statistical Analysis

The experimental data were evaluated for statistical significance using Bonferroni/Dunn’s multiple-range test with GraphPad Prism for Windows, Ver. 5 (GraphPad Software Inc., 2007).

3. Results and Discussion

3.1 Identification of AGEs Formation Inhibitory Active Ingredients of OML-ext

In the preliminary evaluation of mango leaf extract on inhibitory activity of AGEs formation in nonenzymatic glycation of albumin, the OML-ext inhibited AGEs formation with the IC\(_{50}\) value of 43 µg/ml. To identify the active constituents, we carried out activity-guided fractionation of OML-ext using AGEs formation inhibitory assay. Silica gel column chromatographic fractionation of OML-ext gave two active fractions, fraction I and fraction II. The dark green color of fraction I suggested that this fraction may contain green pigments such as chlorophyll as an active ingredient.

Table 1. Inhibitory activities of 3-C-β-D-glucosyl-2,4,4’,6-tetrahydroxybenzophenone (1), mangiferin (2), chlorophyll and cyanidin-3-O-glucoside chloride on AGEs formation

<table>
<thead>
<tr>
<th>Samples</th>
<th>IC(_{50}) values <em>(µM or µg/ml or mM)</em></th>
</tr>
</thead>
<tbody>
<tr>
<td>3-C-β-D-glucosyl-2,4,4’,6-tetrahydroxybenzophenone (1)</td>
<td>85 µM</td>
</tr>
<tr>
<td>Mangiferin (2)</td>
<td>18 µM</td>
</tr>
<tr>
<td>Chlorophyll</td>
<td>41 µg/ml</td>
</tr>
<tr>
<td>Cyanidin-3-O-glucoside chloride</td>
<td>32 µM</td>
</tr>
<tr>
<td>Aminoguanidine hydrochloride</td>
<td>0.9 mM</td>
</tr>
</tbody>
</table>

Aminoguanidine hydrochloride was used as reference compound. a); IC\(_{50}\) value represents the concentration required to inhibit 50% of AGEs
Further purification of another active fraction II led to isolation of 1 and 2 as active constituents. The IC\textsubscript{50} values (Table 1) of 1 and 2 were 85 and 18 \mu M, respectively. As shown in Table 1, the IC\textsubscript{50} value of aminoguanidine hydrochloride as a reference compound was 0.9 mM (= 99 \mu g/ml) in accordance with the reported IC\textsubscript{50} value (138 \mu g/ml) (Shimoda et al., 2011). Thus, a part of the AGES formation inhibitory activity of OML-ext is attributable to these two compounds. To the best of our knowledge, this is the first report on AGES formation inhibitory activity of OML.

As shown in Table 2, OML-ext exhibited an inhibitory activity of AGES formation with the IC\textsubscript{50} value of 43 \mu g/ml. The IC\textsubscript{50} values of young dark reddish brown mango leaf extract (YDL-ext) and young yellow leaf extract (YYL-ext) were 40 and 66 \mu g/ml, respectively (Table 2). The activity of YDL-ext was similar to that of OML-ext. The inhibitory activity of YYL-ext showed slightly decreased compared to those of OML-ext and YDL-ext. HPLC analysis revealed that the contents of 1 and 2 in these leaf extracts were high as described in the preceding paper (Itoh et al., 2016). Taking account of the inhibition data of 1 and 2, and high contents of 1 and 2 in the extracts, the inhibitory activities of these extracts would be partly attributable to these compounds.

Table 2. Inhibitory activities of MeOH extracts of young dark reddish brown and young yellow leaves and old dark green mango leaves on AGES formation

<table>
<thead>
<tr>
<th>Samples</th>
<th>IC\textsubscript{50} values ((\mu g/ml or mM))</th>
</tr>
</thead>
<tbody>
<tr>
<td>Young dark reddish brown leaf extract</td>
<td>40 \mu g/ml</td>
</tr>
<tr>
<td>(=) YDL-ext</td>
<td></td>
</tr>
<tr>
<td>Young yellow leaf extract</td>
<td>66 \mu g/ml</td>
</tr>
<tr>
<td>(=) YYL-ext</td>
<td></td>
</tr>
<tr>
<td>Old dark green leaf extract</td>
<td>43 \mu g/ml</td>
</tr>
<tr>
<td>(=) OML-ext</td>
<td></td>
</tr>
<tr>
<td>Aminoguanidine hydrochloride</td>
<td>0.9 mM</td>
</tr>
</tbody>
</table>

Aminoguanidine hydrochloride was used as reference compound. a); IC\textsubscript{50} value represents the concentration required to inhibit 50\% of AGES formation.

On the other hand, we cannot exclude a hypothesis that other ingredients may also contribute to the activity. Ali et al. (1999) reported that young dark reddish brown mango (cv. Irwin) leaves contain anthocyanin, however, anthocyanin rapidly disappears and chlorophyll content increases with an increase in area after unfolding. Sami & Shakoori (2011) have isolated cyanidin-3-O-glucoside as an anthocyanin with cellulase inhibitory activity from mango leaves. These reports prompted us to evaluate inhibitory effects of chlorophyll and cyanidin-3-O-glucoside on AGES formation, considering with the assumption that the dark green fraction I contained green pigments, such as chlorophyll as one of active ingredients as described above. As shown in Table 1, the IC\textsubscript{50} values of chlorophyll and cyanidin-3-O-glucoside chloride were 41 \mu g/ml, 32 \mu M, respectively. Although AGES formation inhibitory activity of anthocyanins including its inhibitory mechanism has been reported (Chen et al., 2014), there is no report on AGES formation inhibitory effect of chlorophyll. The content of chlorophyll in each leaf extract was spectrophotometrically determined by the method of Porra et al. (1989). The content of total anthocyanin in each leaf extract was determined by a HPLC analysis. As a result, the content of chlorophyll in YDL-ext was 0.85 mg/g. The corresponding content data of other two leaf extracts were as follows; YYL-ext, 2.18 mg/g, and OML-ext, 4.34 mg/g.
with leaves enlargement. The contents of total anthocyanins in these leaf extracts were as follows; YDL-ext, 7.38 ± 0.24 µg/g; YYL-ext, 5.80 ± 0.59 µg/g; OML-ext, not detected of any anthocyanins. These data are in accordance with the reported data of Ali et al. (Ali, Koeda, & Nii, 1999). Considering with the content of chlorophyll and total anthocyanin in leaf extracts, the inhibitory activity of OML-ext was attributable to 1, 2 and 4. The inhibitory activities of YDL-ext and YYL-ext were attributable to 1 and 2, in addition, a part of the inhibitory activity of YDL-ext and YYL-ext was due to anthocyanins whose content is high in young dark reddish brown and young yellow mango leaves. On the other hand, to fully identify other active ingredients and to reveal the inhibitory mechanisms of 1 and chlorophyll, further studies are required, and now undergoing.

From the view point of utility of mango leaves, old dark green leaves obtained by summer pruning may be a reasonable natural resource for the preparation of ingredients with inhibitory activity of AGEs formation.

4. Conclusion

YDL-ext, YYL-ext and OML-ext exhibited inhibitory activities of AGEs formation in nonenzymatic glycation of albumin. The inhibitory activity of YDL-ext was similar to that of OML-ext, and YYL-ext was less potent than YDL-ext and OML-ext, this is the first report to reveal a relationship between leaves maturation and inhibitory activity of AGEs formation. The inhibitory activity of OML-ext was attributable to 3-Cβ-D-glucosyl-2,4,4',6-tetrahydroxybenzophenone (1), mangiferin (2) and chlorophyll. Whereas the inhibitory activity of YDL-ext and YYL-ext were attributable to 1 and 2, in addition, a part of the inhibitory activity of YDL-ext and YYL-ext was due to anthocyanins whose content is high in young dark reddish brown and young yellow mango leaves. This is the first report on AGEs formation inhibitory activity of 1 and chlorophyll. Hitherto, pruned mango leaves were unworthy and discarded during the cultivation process of mango fruit, these findings suggested that pruned mango leaves may be a useful resource for the preparation of ingredients for skin aging or diabetic complications such as diabetic nephropathy with having lipase inhibitory activity. However, further investigations are required to examine administration safety and the mechanisms involved and to reveal other active constituents.

Acknowledgment

We are grateful to all technical staffs of Yuasa Experimental Farm, Kindai University for the collection of mango leaves. I am deeply grateful to Dr. Shunsuke Naruto for his invaluable guidance and advice.

References


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