

In vitro Antioxidant Activity of *Mangifera indica* Leaf Extracts

Kimihisa Itoh¹, Tetsuya Matsukawa^{1,3}, Mamoru Okamoto², Kanasa Minami², Norimichi Tomohiro¹, Kosuke Shimizu¹, Shin'ichiro Kajiyama³, Yuichi Endo^{1,2}, Hideaki Matsuda^{2,4} & Shigeru Shigeoka¹

¹ The Experimental Farm, Kindai University, Wakayama, Japan

² Faculty of Pharmacy, Kindai University, Osaka, Japan

³ Faculty of Biology-Oriented Science and Technology, Kindai University, Wakayama, Japan

⁴ The late professor, Kindai University, Osaka, Japan

Correspondence: Kimihisa Itoh, The Experimental Farm, Kindai University, 2355-2 Yuasa, Yuasa-cho, Arida-gun, Wakayama 643-0004, Japan. Tel: 81-737-622-953. E-mail: itoh_k@nara.kindai.ac.jp

Received: May 14, 2020

Accepted: June 5, 2020

Online Published: June 26, 2020

doi:10.5539/jps.v9n2p39

URL: <https://doi.org/10.5539/jps.v9n2p39>

Abstract

In this study, we aimed to identify the utility of pruned mango (*Mangifera indica* 'Irwin') leaves as a resource for ingredients with antioxidant activity. Firstly, we examined the antioxidant activity of extracts obtained from the pericarps, flesh, flowers, barks, seeds, young dark reddish brown leaves (YDL-ext), young yellow leaves (YYL-ext), and pruned old dark green leaves (OML-ext) obtained from 'Irwin' mango. Among them, methanolic extract of flower and OML-ext showed the most potent 1,1-diphenyl-2-picrylhydrazyl (DPPH) radical scavenging and superoxide dismutase (SOD)-like activity. The flesh extract showed weak DPPH radical scavenging activity, but did not show SOD-like activity. Secondly, we investigated the relationship between the maturation of leaves and their antioxidant activity by considering the contents of their two active polyphenolic components, 3-C- β -D-glucosyl-2,4,4',6-tetrahydroxybenzophenone (**1**) and mangiferin (**2**), in addition to chlorophyll (**3**) and anthocyanins represented by cyanidin-3-O-glucoside (**4**). The DPPH radical scavenging activity of YDL-ext, YYL-ext and OML-ext were mainly attributable to **1**, **2** and **3**, whereas their SOD-like activity was partly attributable to **2**. The DPPH radical scavenging and SOD-like activities of YDL-ext and YYL-ext were attributable to **1** and **2**. These activities were also due to anthocyanins whose content is highest in YDL-ext. Considering the amounts of leaves obtained from pruning, old dark green leaves may be a reasonable natural resource for preparing cosmetics and/or supplemental ingredients with health-enhancing properties, antioxidant activity and inhibitory effect on AGEs formation and pancreatic lipase.

Keywords: anthocyanin, antioxidation, chlorophyll, DPPH radical scavenging activity, 3-C- β -D-glucosyl-2,4,4',6-tetrahydroxybenzophenone, *Mangifera indica* 'Irwin', mangiferin, SOD-like activity

1. Introduction

Mango is one of the most important fruits with more than 1000 cultivars cultivated worldwide. Pruning is an essential component for cultivation of mango fruits. Pruned leaves have hitherto been considered unusable and were thus discarded. Therefore, in the course of our ongoing research to find novel functions of mango leaves, we first focused on the pruned leaves of the mango. We have previously reported that the methanolic extract of pruned old dark green leaves of mango (OML-ext) exhibited inhibitory effects on advanced glycation end products (AGEs) formation and pancreatic lipase (Itoh et al., 2016 and 2017).

Recently, it has been shown that antioxidants play an important role in preventing oxidative stress-related diseases and aging. Numerous data on the antioxidant activities of various plant materials have been reported. In the case of mango, Jose et al. (2018) reported that the ethanolic extracts of mango leaves exhibited antioxidant effects through 1,1-diphenyl-2-picrylhydrazyl (DPPH) radical scavenging and superoxide dismutase (SOD)-like activities with half maximal inhibitory concentration (IC₅₀) values of 14.94 and 68.0 mg/mL, respectively. Another report by Udem, Dahiru, and Etteh (2018) revealed that leaf ethanolic extracts showed the highest percentage of antioxidant activity with 79.09% at 100 mg/mL. According to the report by Fidrianny, Rahmiyani, and Wirasutisna (2013), the ethanolic extract of mango leaf showed potent DPPH radical scavenging activity with IC₅₀ < 50 ppm (\leq 50 μ g/mL). Although, there are several reports concerning the antioxidant activity of mango leaf extract as described above, there is a large discrepancy between the data reported on the potency of

antioxidant activity. If the antioxidant activity of OML-ext is confirmed, the utility of pruned old dark green leaves for the preparing cosmetics and/or supplemental ingredients with health-enhancing properties may be increased.

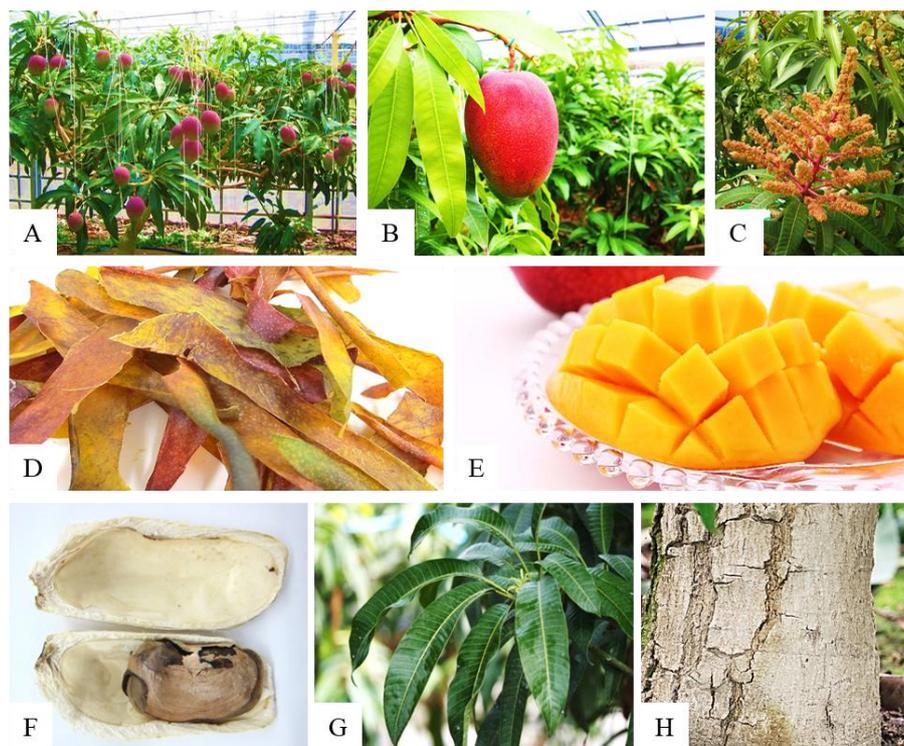
In traditional medicine, the different parts of mango such as seeds, leaves, and bark have been used for their health properties (Lauricella, Emanuele, Calvaruso, Giuliano, & D'Anneo, 2017).

Firstly, these interesting bibliographies have inspired us to examine the antioxidant activity of the methanolic extracts of several parts such as pericarps, flesh, flowers, bark, seeds, from the mango cultivar 'Irwin' cultivated in the Experimental Farm of Kindai University. Since mango is an evergreen plant, leaves at different three stages of growth, young dark reddish brown leaves, young yellow leaves and pruned old dark green leaves, were collectable during pruning at the same time in late summer. This prompted us to examine the relationship between the maturation of leaves and these antioxidant activities. To the best of our knowledge, there has been no investigation of the relationship between mango leaves maturation and their antioxidant activities by considering the content of their four chemical components, two polyphenolic components, 3-C- β -D-glucosyl-2,4,4',6-tetrahydroxybenzophenone (1) and mangiferin (2), in addition to chlorophyll (3) and anthocyanins represented by cyanidin-3-O-glucoside (4).

2. Materials and Method

2.1 Plant Materials

The pericarps, flesh, flowers, bark, seeds, and pruned old dark green leaves of *Mangifera indica* 'Irwin' were collected from the Experimental Farm of Kindai University (34° 2' N, 135° 11' E, 17 m ASL), located in Wakayama Prefecture, Japan (Figure 1). The *M. indica* trees planted in the ground are commercially grown in a plastic greenhouse under controlled conditions (temperature: winter, min. 2 °C (room) and 10 °C (soil); summer, max. 35 °C (room) and 31 °C (soil)). 'Irwin' mango is a representative cultivar especially in Japan and Taiwan because of its early ripening and relatively good resistance to cold weather.



The picture of A, Irwin mango tree with several fruits in a greenhouse, Kindai University; B, fruit; C, flowers; D, pericarps; E, flesh; F, seed; G, old dark green leaves; H, bark.

Figure 1. Fruit, flowers, pericarps, flesh, seed, old dark green leaves and bark of 'Irwin' mango prepared in the experiment

Three kinds of mango leaves (old dark green leaf, young dark reddish brown leaf, and young yellow leaf) were collected from 250 mango trees which were propagated by grafting (height of trees: 2 m, age of trees: 18–26 years old, life span of trees: 40–50 years). The materials (old dark green leaves, young dark reddish brown leaves, and young yellow leaves) were collected in August 2015 and August 2016. These young and old leaves were collected together at the time of pruning in late summer which was possible as mango is an evergreen plant. The collected leaves were visually classified by color into three groups, namely young dark reddish brown, young yellow, and old dark green leaves as illustrated in our previous papers (Itoh et al., 2016 and 2017). The samples were identified by the Experimental Farm of Kindai University, air-dried at 50 °C for 72 h in an automatic air-drying apparatus (Vianove Inc., Tokyo, Japan), and powdered. Voucher specimens of pericarps, flesh, flowers, bark, seeds and leaves (Mango old leaf, OML201508DG-S and OML201608DG-S; Mango young dark reddish brown leaf, YML201508DB-S; Mango young yellow leaf, YML201508Y-S) are deposited at the Experimental Farm, Kindai University.

2.2 Extraction

Methanolic extracts of young dark reddish brown leaves (YDL-ext), young yellow leaves (YYL-ext), and old dark green leaves (OML-ext) were obtained according to the previous paper (Itoh et al., 2016). The yields of the methanolic extract obtained from pericarps, flesh, flowers, bark and seeds of *M. indica* 'Irwin' were 23%, 63%, 25%, 19%, and 6%, respectively.

2.3 Reagents

The chemical and biochemical reagents used in this study were of reagent grade and were purchased from Wako Pure Chemical Industries, Ltd. (Osaka, Japan) and/or Nacalai Tesque, Inc. (Kyoto, Japan) unless otherwise noted.

2.4 In vitro Radical Scavenging Activity

Radical scavenging activity was measured according to the method outlined by Blois (1958) with minor modifications described in a previous paper (Itoh et al., 2009). Briefly, the test sample was dissolved in DMSO and diluted with 0.5 M acetate buffer (pH 5.5) to a final DMSO concentration of 5% v/v. A mixture of test sample solution (2 mL), EtOH (1.6 mL), 0.5 M acetate buffer (0.4 mL), and 0.5 mM DPPH/EtOH solution (1.0 mL) was allowed to stand for 30 min at room temperature. The OD of the resulting mixture was determined with a spectrophotometer at 520 nm. L-Ascorbic acid was used as a reference agent. The scavenging activity of each sample was expressed as the percentage of the decrease in OD compared with that of control DPPH solution. The IC₅₀ value represents the concentration of sample required to scavenge 50% of DPPH free radicals. Each assay was performed in triplicate (*P*-value < 0.01).

2.5 In vitro SOD-Like Activity

SOD-like activity was measured according to the method outlined by Oyanagui (1984) with minor modifications described in a previous paper (Itoh et al., 2009). Briefly, the test sample was dissolved in DMSO and diluted with 0.5 mM disodium dihydrogen ethylenediamine tetraacetate (EDTA)-PBS buffer (pH 8.2) to a final DMSO concentration of 1% v/v. SOD, as a reference, originating from cow's milk (Roche Co., Tokyo, Japan), was dissolved with the buffer. A mixture of the buffer (0.2 mL), 0.5 mM hypoxanthine in the buffer (0.2 mL), reagent A solution (10 mM hydroxylamine hydrochloride and 1 mg/mL hydroxylamine-*o*-sulfonic acid in water) (0.1 mL), water (0.2 mL), and the sample solution (0.1 mL) was preincubated at 37 °C for 10 min. Five mU/mL xanthine oxidase (Roche Co., Tokyo, Japan) solution in the buffer (0.2 mL) was added to the above solution, and the mixture was incubated at 37 °C for 30 min. Reagent B (30 μM *N*-1-naphthylethylenediamine · 2HCl, 3 mM sulfanilic acid, and 25% acetic acid in water) (2 mL) was added to the reaction mixture. The resulting mixture was allowed to stand for 30 min at room temperature, and then OD was measured at 550 nm with a spectrophotometer. The SOD-like activity of each sample was expressed as the percentage of the decrease in OD compared with that of control A or B solution. The IC₅₀ value represents the concentration of sample required to scavenge 50% of the superoxide anions produced by the hypoxanthine-xanthine oxidase system. Each assay was performed in triplicate (*P*-value < 0.01).

2.6 Determination of Content of 1, 2, 3 and 4 in Each Mango Cultivar Leaf Extract

In this paper, determination of each compound in the leaf extracts was performed in triplicate, and the values are represented as the mean ± standard deviation. The HPLC determination method for the contents (mg/g extract) of **1** and **2** in each mango leaf extract was described in a previous paper (Itoh et al., 2016). Spectrophotometric determination of the content of **3** (mg/g extract) was carried out according to the method given by Porra, Thompson, and Kriedemann (1989). The content of total anthocyanin (μg/g extract) in each leaf extract was

determined by the HPLC analysis described in a previous paper (Itoh et al., 2017).

2.7 Statistical Analysis

The experimental data were evaluated for statistical significance using Bonferroni/Dunn's multiple-range test with GraphPad Prism for Windows, Ver. 5 (GraphPad Software Inc., 2007; Armonk, NY, USA).

3. Results and Discussion

3.1 Antioxidant Activity of the Extracts Obtained from Pericarps, Flesh, Flowers, Bark, Seeds and Pruned Old Dark Green Leaves (OML-ext)

We examined the *in vitro* antioxidant activity of the extracts obtained from several parts of 'Irwin' mango such as pericarps, flesh, flowers, bark, seeds and pruned old dark green leaves by two assays that estimated the DPPH radical scavenging and SOD-like activities. 'Irwin' mangoes are commercially grown in a plastic greenhouse, and all parts of mango were collected from the Experimental Farm of Kindai University (Photo 1) for preparation of each extract. Among them, flower extract and OML-ext showed more potent DPPH radical scavenging and SOD-like activities than the other extracts as shown in Table 1. The DPPH radical scavenging activity of flesh extract was weak, but the extract did not show SOD-like activity. The scavenging activity of seed extract against DPPH radical was active, whereas the SOD-like activity was very weak. Moreover, the SOD-like activities of pericarps extract and bark extract were also weak. The DPPH radical scavenging activity assay method has been widely used to evaluate the radical scavenging activity of plant extracts and their constituents, and the IC₅₀ value represents the concentration of sample required to scavenge for 50% of DPPH free radicals (Hou et al., 2003). The scavenging activity was evaluated using L-ascorbic acid (IC₅₀, 22 µM), a reference agent, whose IC₅₀ was in accordance with our reported IC₅₀ value (30 µM) (Itoh et al., 2009). In SOD-like activity assay, SOD was used as a reference agent, the IC₅₀ value of SOD was 0.2 U/mL in accordance with our reported IC₅₀ value (0.2 U/mL) (Itoh et al., 2009). Considering the potency of the activity of leaf extract in preliminary examination, and the amounts of leaves obtained from pruning, old dark green leaves may be a reasonable natural resource for the preparation of ingredients with antioxidant activity.

Table 1. IC₅₀ values of DPPH radical scavenging and SOD-like activities of MeOH extracts obtained from 'Irwin' mango leaves, flowers, pericarps, seeds, bark and flesh

Samples	DPPH radical scavenging activity ^{a)} (µg/mL or µM)	SOD-like activity ^{b)} (µg/mL or U/mL)
Leaf extract (OML-ext)	9 µg/mL	117 µg/mL
Flower extract	3 µg/mL	171 µg/mL
Pericarp extract	11 µg/mL	760 µg/mL
Seed extract	4 µg/mL	888 µg/mL
Bark extract	12 µg/mL	656 µg/mL
Flesh extract	768 µg/mL	N.E. ^{c)}
L-Ascorbic acid	22 µM	N.D. ^{d)}
SOD	N.D. ^{d)}	0.2 U/mL

^{a)} IC₅₀ value represents the concentration of sample required to inhibit 50% of DPPH free radical; ^{b)} IC₅₀ value represents the concentration of sample required to inhibit 50% of superoxide anions; ^{c)} N.E.: no effect; ^{d)} N.D.: not determined.

3.2 Relationship between Leaves Maturation and Their Antioxidant Activities

We investigated the relationship between leaves maturation and their antioxidant activities by considering the content of their two active polyphenolic components, 3-C-β-D-glucosyl-2,4,4',6-tetrahydroxybenzophenone (**1**) and mangiferin (**2**), in addition to the content of chlorophyll (**3**) and anthocyanins represented by cyanidin-3-O-glucoside (**4**). The antioxidant activities of these four major components of mango leaves were examined and the results were depicted in Table 2. The collected leaves were visually classified by color into three groups as described in the previous paper (Itoh et al., 2017). The antioxidant activity of the extract of three stage of leaves maturation of mango leaves (young dark reddish brown leaves (YDL-ext), young yellow leaves (YYL-ext) and old dark green leaves (OML-ext)) is shown in Table 3. The antioxidant activity of OML-ext was superior to those of YDL-ext and YYL-ext, and the activities of these extracts were slightly increased when the leaves matured from yellow to dark green. As for the active ingredients that reportedly inhibit AGEs formation,

the activity-guided fractionation of OML-ext using these assays and HPLC analysis led to the isolation of **1**, **2** and **3** as one of the active constituents of OML-ext, and **1**, **2** and **4** as one of the active constituents of YDL-ext and YYL-ext (Itoh et al., 2017; Itoh et al., 2020). Moreover, further HPLC analysis revealed that the contents (mg/g or µg/g extract) of **1**, **2**, **3** and **4** in these leaf extracts were high as shown in Table 4. As a result, the DPPH radical scavenging activity of YDL-ext, YYL-ext and OML-ext was partly attributable to **1**, **2** and **3**, and their SOD-like activity was partly attributable to **2**. The DPPH radical scavenging and SOD-like activities of YDL-ext and YYL-ext were due to anthocyanins represented by **4** whose content is higher in young mango leaves. Considering the potent activity of leaf extract in preliminary examination, and the amounts of leaves obtained from pruning, old dark green leaves may be a reasonable natural resource for preparing ingredients with antioxidant activity. These findings identified the utility of the old dark green mango leaves considering with the potent activity of leaf extract in preliminary examination, and with the amounts of leaves obtained from pruning, thus the pruned old dark green mango leaves may be a reasonable natural resource for preparing ingredients with antioxidant activity.

Malherbe et al. (2014) have reported that no radical scavenging was observed for **1**, when isolated from shoots of *Cyclopia genistoides*, using the on-line HPLC-DPPH assay, whereas **2** had high activity against DPPH radical in our experiments. Because they did not use a reference compound such as L-ascorbic acid or α-tocopherol, it is difficult to discuss the reasons here. We assumed that the discrepancy of the activity of **1** might be partially due to some differences in experimental conditions, such as evaluation method. Accordingly, this is the first report on DPPH radical scavenging and SOD-like activities of **1**. On the other hand, the potent inhibitory activity of leaf extracts cannot exclude a hypothesis that other ingredients may also contribute to the activity. To identify other active ingredients, further studies are required, which are now ongoing.

Table 2. IC₅₀ values of DPPH radical scavenging and SOD-like activities of 3-C-β-D-glucosyl-2,4,4',6-tetrahydroxybenzophenone (**1**), mangiferin (**2**), chlorophyll (**3**) and cyanidin-3-O-glucoside (**4**)

Samples	DPPH radical scavenging activity ^{a)} (µM or µg/mL)	SOD-like activity ^{b)} (µM or U/mL)
1	74 µM	N.E. ^{c)}
2	12 µM	872 µM
3	224 µg/mL	N.E. ^{c)}
4	6 µM	196 µM
L-Ascorbic acid	22 µM	N.D. ^{d)}
SOD	N.D. ^{d)}	0.2 U/mL

^{a)} IC₅₀ value represents the concentration of sample required to inhibit 50% of DPPH free radical; ^{b)} IC₅₀ value represents the concentration of sample required to inhibit 50% of superoxide anions. ^{c)} N.E.: no effect, ^{d)} N.D.: not determined.

Table 3. IC₅₀ values of DPPH radical scavenging and SOD-like activities of MeOH extracts of young dark reddish brown, young yellow and old dark green mango leaves

Samples	DPPH radical scavenging activity ^{a)} (µg/mL or µM)	SOD-like activity ^{b)} (µg/mL or U/mL)
Young dark reddish brown leaf extract (YDL-ext)	29 µg/mL	141 µg/mL
Young yellow leaf extract (YYL-ext)	15 µg/mL	138 µg/mL
Old dark green leaf extract (OML-ext)	9 µg/mL	117 µg/mL
L-Ascorbic acid	22 µM	N.D. ^{c)}
SOD	N.D. ^{c)}	0.2 U/mL

^{a)} IC₅₀ value represents the concentration of sample required to inhibit 50% of DPPH free radical; ^{b)} IC₅₀ value represents the concentration of sample required to inhibit 50% of superoxide anions. ^{c)} N.D.: not determined.

Table 4. The contents of 3-C- β -D-glucosyl-2,4,4',6-tetrahydroxybenzophenone (**1**), mangiferin (**2**), chlorophyll (**3**) and cyanidin-3-O-glucoside (**4**) in YDL-ext, YYL-ext and OML-ext

Samples	1 (mg/g extract)	2 (mg/g extract)	3 (mg/g extract)	4 (μ g/g extract)
YDL-ext	400.0 \pm 11.0 mg/g	78.6 \pm 1.2 mg/g	0.85 \pm 0.002 mg/g	7.38 \pm 0.24 μ g/g
YYL-ext	278.6 \pm 15.5 mg/g	79.4 \pm 1.0 mg/g	2.18 \pm 0.004 mg/g	5.80 \pm 0.59 μ g/g
OML-ext	205.9 \pm 7.6 mg/g	85.1 \pm 0.5 mg/g	4.34 \pm 0.017 mg/g	N.D. ^{a)}

YDL-ext: young dark reddish brown leaf extract, YYL-ext: young yellow leaf extract, OML-ext: old dark green leaf extract. ^{a)} N.D.: not detected.

4. Conclusion

The antioxidant activity of the extracts obtained from several parts of pericarps, flesh, flowers, bark, seeds and old dark green leaves obtained from *Mangifera indica* 'Irwin' was examined by evaluating DPPH radical scavenging and SOD-like activities. Among them, flower extract and OML-ext showed more potent DPPH radical scavenging and SOD-like activities than those of other extracts. Considering the potency of the activity of leaf extract in preliminary examination, and the amounts of leaves obtained from pruning, old dark green leaves may be a reasonable natural resource for the preparation of ingredients with antioxidant activity. Among the comparison in the activities between YDL-ext, YYL-ext and OML-ext, both activities of OML-ext were superior to those of YDL-ext and YYL-ext, while the activities of these extracts were slightly increased in accordance with the maturation of the leaves from yellow to dark green. The DPPH radical scavenging activity of YDL-ext, YYL-ext and OML-ext was partly attributable to **1**, **2** and **3**, and their SOD-like activity was partly attributable to **2**. The DPPH radical scavenging and SOD-like activities of YDL-ext and YYL-ext might be partially due to the higher anthocyanin content in young mango leaves. This is the first report on DPPH radical scavenging and SOD-like activities of **1**. Hitherto, pruned mango leaves were unworthy and discarded during the cultivation process of mango fruits. Thus, from the viewpoint of utility of mango leaves, the pruned old dark green leaves may be a reasonable natural resource for the preparation of cosmetics and/or supplemental ingredients with health-enhancing properties and antioxidant activity in addition to the known inhibitory effects on AGEs formation (Itoh et al., 2017) and pancreatic lipase (Itoh et al., 2016). Confirming the antioxidant activity of mango leaf increased the utility of the pruned mango leaves. However, further investigations are required to examine the safety of administration and the mechanisms involved and also to reveal other active constituents.

Acknowledgments

We are grateful to the technical staff of Yuasa Experimental Farm, Kindai University for the collection of mango leaves. We are also deeply grateful to Dr. Shunsuke Naruto for his invaluable guidance and advice. We would like to thank Editage (www.Editage.com) for English language editing.

References

- Blois, M. S. (1958). Antioxidant determinations by the use of a stable free radical. *Nature*, *181*, 1199-1200. <https://doi.org/10.1038/1811199a0>
- Fidrianny, I., Rahmiyani, I., & Wirasutisna, R. K. (2013). Antioxidant capacities from various leaves extracts of four varieties mangoes using DPPH, ABTS assays and correlation with total phenolic, flavonoid, carotenoid. *International Journal of Pharmacy and Pharmaceutical Sciences*, *5*(4), 189-194.
- Hou, W. C., Lin, R. D., Cheng, K. T., Hung, Y. T., Cho, C. H., Chen, C. H., ... Lee, M. H. (2003). Free radical-scavenging activity of Taiwanese native plants. *Phytomedicine*, *10*, 170-175. <https://doi.org/10.1078/094471103321659898>
- Itoh, K., Hirata, N., Masuda, M., Naruto, S., Murata, K., Wakabayashi, K., & Matsuda, H. (2009). Inhibitory effects of *Citrus hassaku* extract and its flavanone glycosides on melanogenesis. *Biological and Pharmaceutical Bulletin*, *32*(3), 410-415. <https://doi.org/10.1248/bpb.32.410>
- Itoh, K., Murata, K., Nakagaki, Y., Shimizu, A., Takata, Y., Shimizu, K., ... Matsuda, H. (2016). Inhibitory activity of *Mangifera indica* leaf extract on pancreatic lipase. *Journal of Plant Studies*, *5*(2), 72-78. <https://doi.org/10.5539/jps.v5n2p72>
- Itoh, K., Murata, K., Sakaguchi, N., Akai, K., Yamaji, T., Shimizu, K., ... Matsuda, H. (2017). Inhibition of advanced glycation end products formation by *Mangifera indica* leaf extract. *Journal of Plant Studies*, *6*(2), 102-107. <https://doi.org/10.5539/jps.v6n2p102>

- Itoh, K., Matsukawa T., Minami K., Okamoto M., Tomohiro N., Shimizu K., ... Shigeoka S. (2020). Inhibitory effect of several *Mangifera indica* cultivar leaf extracts on the formation of advanced glycation end products (AGEs). *Journal of Plant Studies*, 9(2), 33-38. <https://doi.org/doi:10.5539/jps.v9n2p33>
- Jose, M., Varghese, I. V., Jayakar, V., Lokapur, V., Srinivasa, K., & Shantaram, M. (2018). Evaluation of antioxidant activities and free radical scavenging properties in mango leaves, husks of areca and coconut. *Research Journal of Pharmaceutical, Biological and Chemical Sciences*, 9(4), 1607-1619.
- Lauricella, M., Emanuele, S., Calvaruso, G., Giuliano, M., & D'Anneo, A. (2017). Multifaceted health benefits of *Mangifera indica* L. (Mango): The inestimable value of orchards recently planted in Sicilian rural areas. *Nutrients*, 9(5), 525-538. <https://doi.org/10.3390/nu9050525>
- Malherbe, C. J., Willenburg, E., de Beer, D., Bonnet, S. L., van der Westhuizen, J. H., & Joubert, E. (2014). Iriflophenone-3-C-glucoside from *Cyclopia genistoides*: Isolation and quantitative comparison of antioxidant capacity with mangiferin and isomangiferin using on-line HPLC antioxidant assays. *Journal of Chromatography B*, 951-952, 164-171. <https://doi.org/10.1016/j.jchromb.2014.01.038>
- Oyanagui, Y. (1984). Reevaluation of assay methods and establishment of kit for superoxide dismutase activity. *Analytical Biochemistry*, 142(2), 290-296. [https://doi.org/10.1016/0003-2697\(84\)90467-6](https://doi.org/10.1016/0003-2697(84)90467-6)
- Porra, R. J., Thompson, W. A., & Kriedemann, P. E. (1989). Determination of accurate extinction coefficients and simultaneous equations for assaying chlorophylls *a* and *b* extracted with four different solvents: verification of the concentration of chlorophyll standards by atomic absorption spectroscopy. *Biochimica et Biophysica Acta (BBA) - Bioenergetics*, 975(3), 384-394. [https://doi.org/10.1016/S0005-2728\(89\)80347-0](https://doi.org/10.1016/S0005-2728(89)80347-0)
- Udem, C. G., Dahiru, D., & Etteh, C. C. (2018). *In vitro* antioxidant activities of aqueous and ethanol extracts of *Mangifera indica* leaf, stem-bark and root-bark. *Pharmacognosy Communications*, 8(3), 119-124. <http://dx.doi.org/10.5530/pc.2018.3.25>

Copyrights

Copyright for this article is retained by the author(s), with first publication rights granted to the journal.

This is an open-access article distributed under the terms and conditions of the Creative Commons Attribution license (<http://creativecommons.org/licenses/by/4.0/>).