Pre-Plant Soaking of Corms in Growth Regulators Influences the Multiple Sprouting, Floral and Corm Associated Traits in *Gladiolus grandiflorus* L.

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Abstract

*Gladiolus* is an ornamental bulbous plant and is commercially propagated in the world through its corms. Usually, a single bud is sprouted from a mother corm but sprouting of multiple buds favors an increase in production of propagules. Thus, present research work was carried out to enhance the sprouting of multiple buds and evaluate its effects on other growth parameters through pre-plant soaking of corms in solution of plant growth regulators. The corms were soaked in solutions of gibberellic acid (GA$_3$), benzyladenine (BA) or ethrel at 0, 50, 100 or 150 ppm concentrations for 24 hrs before planting in the field. Gibberellic acid at 100ppm concentration increased plant height to 105 cm compared to 97.60 cm in control plants, flowering percentage (84.67%), spike length (40.03 cm) and also boosted the corm weight (68.30 g). Soaking of corms in 150 ppm benzyladenine solution enhanced the number of sprouting per corm (2.14) and reduced the plant height (87.00 cm) while 50 ppm concentration of ethrel increased the spike length (42.14 cm). In conclusion, soaking of corms in benzyladenine favored the modifications in various traits of interest including sprouting of multiple buds and an increase in the production of corms while gibberellic acid improved the floral characteristics of gladiolus spikes.

Keywords: corm treatment, corm multiplication, gladiolus, plant growth regulators, production system

1. Introduction

*Gladiolus* is a perennial bulbous plant that belongs to family Iridaceae and widely grown as a cut flower in the world. The genus “*Gladiolus*” consists of 260 species, out of which 250 belong to sub-Saharan Africa while 10 species from Eurasia (Goldblatt & Manning, 1998; Manning & Goldblatt, 2008). The gladiolus is commercially propagated through its corm which is an underground modified stem that provides nutrients during sprouting (Ghamsari et al., 2007).

The conventional production system of gladiolus allows to get usually one sprout and one daughter corm from a mother corm but sprouting of more buds from a single corm favors to increase in production of corms. The minimum control over vegetative and floral traits is another drawback of conventional system, which is not in favor of meeting the market demands. The control over flowering time and floral characteristics according to the demand of market has been achieved in many cut flowers by adopting modern production techniques including the use of plant growth regulators (PGRs). The application of PGRs has become the part of their cultural practices in many ornamental plants to modify their vegetative and floral traits. There are different application methods of PGRs including foliar application, pre-plant soaking and drenching but foliar application is the most common method being practiced in ornamental plants. Soaking of bulbs in solution of plant growth regulators has been used (Larson et al., 1987) and it is an efficient method for obtaining good results and also have advantages over other methods in terms of time, labor saving and accurate dosage (Ranwala et al., 2002). Keeping in view the efficiency of this method, Pre-plant dipping of planting material in a chemical solution is...
now becoming a popular method among commercial growers (Schnelle et al., 2005).

PGRs consist of numerous compounds including gibberellins, auxins, cytokinins, abscisic acid and ethylene. The use of plant growth regulators is a successful approach for improvement of various traits, including, number of flower buds (Boyle, 1992), induction of early flowering (Sakai et al., 1979), delay flowering (Paulin & Muloway, 1979), flower development (Irish, 2009) and increase yield (Malabug et al., 2010). PGRs has been used in the commercial production of ornamental plants (Rademacher, 1991) but their efficacy depends on different plant species (Dole & Wilkins, 1999).

Gibberellic acid has an important role in different plant processes, including seed germination, stem elongation, leaf expansion and flower development (Olszewski et al., 2002). Ethrel is a chemical compound containing ethephon as an active ingredient, releases ethylene which act as plant growth regulators. Application of ethrel is effective in modifying different attributes of ornamental plants. The cytokinins are an important part of several commercial plant growth regulators (PGRs) being used on different plant. Application of Cytokinins has been reported to stimulate the growth of axillary buds in plants. Benzyladenine is also a cytokinin and its application is reported in different ornamental bulbous plants including tulips (de Munk & Gijzenberg, 1977), lilies (Ranwala & Miller, 1998) and calla bulbs (Naor et al., 2005).

The objective of this study was to evaluate the effects of pre-plant application of PGRs on sprouting of multiple buds, production of corms, cormels, floral traits, and also to find out interaction among these traits, that can help to improve the conventional production system of gladiolus.

2. Materials and Methods

2.1 Plant Material and Treatments

The healthy corms of gladiolus cultivar “Amsterdam” were purchased from the local market of Lahore, Pakistan. The corms were dipped in a freshly prepared solution of gibberellic acid, benzyladenine or ethrel at different concentrations of 50, 100 or 150 ppm for 24 hrs. The control corms were dipped in distilled water for the same time. The treated and non treated corms were planted in the field of floriculture research area, University of Agriculture, Faisalabad, Pakistan, on ridges at the plant to plant distance of 15 cm while row to row distance was 60 cm. The field area is located at 31°26′N latitude, 73°06′E longitude and 184.4 m altitude. All the cultural practice including irrigation, weeding, fertilizers were same for all plants. The irrigation was scheduled at the interval of 7-10 days according to the condition of soil while NPK fertilizer was applied at 100, 90, 100 kg/acre, respectively and weeding was done manually. The experiment was conducted in randomized complete block design (RCBD) with three replicates having 60 corms in each treatment.

2.2 Data Collection

The plants were allowed to grow in the field and data was recorded according to the growth stage of plants. The data for different vegetative variables, including days to sprouting, number of buds sprouted per corm, vegetative plant height (cm) was recorded before the initiation of flowering while data for total plant height (cm), days to open 1st floret, spike length (cm), spike diameter (cm), 1st floret weight (g), 1st floret diameter (cm) and flowering percentage was noted after the opening of florets in the spike. Lifting of corms from the field was done after 70 days of flowering, and the data for corm related attributes including, corm diameter (cm), corm weight (g), number of corms per plant, total number of cormels per plant, total cormel weight per plant (g) and average weight of cormel (g) was recorded.

2.3 Statistical Analysis

The recorded data was subjected to analysis of variance and treatment means were compared by applying Duncan’S Multiple Range (DMR) test by using SAAS software version 1.101. Pearson correlation among different variables was found by using SPSS ver. 16 software.

3. Results

3.1 Vegetative Variables

Corms treated with gibberellic acid showed 100% sprouting within 10 days of sowing compared to 75% sprouting in control (Figure 1). Benzyladenine delayed the sprouting process and 56% sprouting recorded in 10 days. All the treatment showed 100% sprouting but after 15 days of sowing. The significant effect of treatments on the number of sprouted buds per corm was shown in Table 1. The control corms showed only 1.02 sprouts per corm while benzyladenine treatment at 150 ppm increased the sprouting of buds to 2.14 followed by 1.69 at 100 ppm. Foliage plant height also increased by the application of 100 ppm ethrel to 78.26 cm compared to 70.18 cm in control plants. Gibberellic acid at 100 ppm concentration also increased the foliage plant height (75.26 cm).
The soaking of corms in gibberellic acid at 100 ppm concentration significantly increased the total height of gladiolus plants to 105.80 cm compared to 97.60 cm in control plants. The maximum plant height (110.98 cm) was recorded in the 100 ppm concentration of ethrel treatment.

![Figure 1. Effect of growth regulators on sprouting percentage of gladiolus corms](image)

3.2 Floral Variables

Application of plant growth regulators also changed the flower characteristics of gladiolus spikes. Gibberellic acid at 100 ppm concentration significantly decreased the days (70.88) to opening of the first floret on the spike followed by 70.97 days in 50 ppm of ethrel (Table 2). Benzyladenine caused the significant delay in flowering and its 150 ppm application increased days to opening of the first floret to 107.50 days compared to 77.05 days in control plants. Flowering percentage increased to 84.67% in the treatment of 100 ppm gibberellic acid compared to 71.67% in control plants. Benzyladenine also increased the flowering percentage (80.33%) in 50 ppm treatment, but decreased in comparatively higher concentrations and minimum flowering percentage (21%) was recorded in the 150 ppm dose.

![Figure 1. Effect of growth regulators on sprouting percentage of gladiolus corms](image)

### Table 1. Effect of growth regulators on vegetative variables of gladiolus

<table>
<thead>
<tr>
<th>Treatments (ppm)</th>
<th>Sprouted buds per corm</th>
<th>Foliation Plant Height (cm)</th>
<th>Total Plant Height (cm)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>1.02 ± 0.02 e</td>
<td>70.18 ± 0.76 d</td>
<td>97.60 ± 1.50 cd</td>
</tr>
<tr>
<td>GA3 50</td>
<td>1.06 ± 0.04 de</td>
<td>71.06 ± 0.04 d</td>
<td>99.63 ± 1.95 c</td>
</tr>
<tr>
<td>GA3 100</td>
<td>1.29 ± 0.08 cde</td>
<td>75.26 ± 1.67 bc</td>
<td>105.80 ± 0.35 b</td>
</tr>
<tr>
<td>GA3 150</td>
<td>1.25 ± 0.14 cde</td>
<td>74.58 ± 1.49 bc</td>
<td>105.47 ± 0.65 b</td>
</tr>
<tr>
<td>BA 50</td>
<td>1.44 ± 0.04 bc</td>
<td>72.38 ± 0.22 cd</td>
<td>100.79 ± 0.70 c</td>
</tr>
<tr>
<td>BA 100</td>
<td>1.69 ± 0.04 b</td>
<td>70.00 ± 1.48 d</td>
<td>93.13 ± 2.24 e</td>
</tr>
<tr>
<td>BA 150</td>
<td>2.14 ± 0.08 a</td>
<td>64.20 ± 0.55 e</td>
<td>87.00 ± 0.58 f</td>
</tr>
<tr>
<td>Ethrel 50</td>
<td>1.13 ± 0.07 de</td>
<td>76.90 ± 2.01 ab</td>
<td>110.98 ± 2.15 a</td>
</tr>
<tr>
<td>Ethrel 100</td>
<td>1.32 ± 0.22 cd</td>
<td>78.26 ± 0.84 a</td>
<td>109.20 ± 0.35 ab</td>
</tr>
<tr>
<td>Ethrel 150</td>
<td>1.01 ± 0.01 e</td>
<td>70.90 ± 1.67 d</td>
<td>94.25 ± 0.72 de</td>
</tr>
</tbody>
</table>

Note. GA3: Gibberellic acid; BA: Benzyladenine. Means sharing the same letter do not differ by DMR test at 5% probability level; ± means SE.

Control plants showed spike length of 34.75 cm and maximum increase (42.14 cm) was recorded in 50 ppm treatment of ethrel. Spike diameter was also increased to 0.86 cm in 50 ppm ethrel compared to 0.75 cm in control plants. Application of 100 ppm ethrel showed a maximum increase (12.88) in a number of florets per spike while the minimum increase (9.90) was recorded in 150 ppm ethrel. The maximum increase in floret...
diameter (8.98 cm) was exhibited in the 150 ppm concentration of gibberellic acid while benzyladenine at same concentration showed a minimum increase (6.31 cm) in floret diameter. The 1st floret weight was increased to 5.27 g in application of 150 ppm gibberellic acid compared to 4.27 g in control plants.

3.3 Corms Variables

The treatments significantly affected the different attributes of corms and cormels (Table 3). Benzyladenine at 150 ppm concentration showed a maximum increase (2.50) in the number of corms per plant while the minimum increase in corm diameter (3.90 cm) was also recorded in this treatment. Benzyladenine induced the multiple shoot induction which also resulted in production of multiple daughter corms from a single parent corm as shown in figure 2. Application of gibberellic acid at 100 ppm caused the maximum increase in corm diameter (6.01 cm) compared to 5.81 cm in control plants. Corm weight was significantly increased (68.30 g) in response to application of 100 ppm gibberellic acid followed by 66.51 g in the 100 ppm concentration of ethrel. The highest increase in the number of cormels per plant (31.33) was observed in gibberellic acid treatment at 150 ppm concentration while 8.57 cormels per plant recorded in control plants. Total cormel weight per plant was increased to 5.96 g in 150 ppm concentration of ethrel while control plants gave the maximum increase in average cormel weight (0.41 g).

3.4 Correlation among Variables

Number of sprouted buds per corm showed significant negative correlation with the other vegetative, floral and corm related parameters except the corms per plant (Table 4). Sprouting of buds per corm is significantly affected by the application of PGRs especially the benzyladenine. This parameter greatly influenced on altering the other valuable parameters and found as key to control the plant growth in the present research. The second most influential parameter was days to flower which showed negative correlation with the floral and corm associated parameters except the corms per plant which is positively correlated. On the other hand, vegetative plant height showed significantly positive correlation with all parameters of floral and corm growth except days to flower and corms per plant which are negatively correlated. Parameters associated with cormels are not significantly correlated with the vegetative and floral variables, but the number of cormel per plant showed significant correlation with other parameters of cormels including total cormel weight and average cormel weight. It means that the total weight of cormels increased with the increase in the total number of cormels but the average weight of a cormel decreased.

Table 2. Effects of growth regulators on floral variables of gladiolus

<table>
<thead>
<tr>
<th>Treatments (ppm)</th>
<th>Days to flower</th>
<th>Flowering %</th>
<th>Spike length (cm)</th>
<th>Spike diameter (cm)</th>
<th>Florets per spike</th>
<th>Floret diameter (cm)</th>
<th>Floret weight (g)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>77.05 ± 1.10 cde</td>
<td>71.67 ± 3.53 d</td>
<td>34.57 ± 0.82 cd</td>
<td>0.75 ± 0.01 cd</td>
<td>10.24 ± 0.05 de</td>
<td>7.95 ± 0.19 c</td>
<td>4.27 ± 0.45 cd</td>
</tr>
<tr>
<td>GA3 50</td>
<td>73.80 ± 1.15 def</td>
<td>72.67 ± 2.33 d</td>
<td>37.85 ± 0.95 bc</td>
<td>0.78 ± 0.01 b</td>
<td>11.25 ± 0.14 bc</td>
<td>8.27 ± 0.13 bc</td>
<td>4.31 ± 0.49 c</td>
</tr>
<tr>
<td>GA3 100</td>
<td>70.88 ± 0.65 f</td>
<td>84.67 ± 2.40 a</td>
<td>40.03 ± 0.79 ab</td>
<td>0.79 ± 0.02 b</td>
<td>12.78 ± 0.01 a</td>
<td>8.94 ± 0.41 a</td>
<td>5.23 ± 0.42 a</td>
</tr>
<tr>
<td>GA3 150</td>
<td>71.85 ± 0.20 ef</td>
<td>83.33 ± 2.73 ab</td>
<td>37.69 ± 0.01 bc</td>
<td>0.76 ± 0.04 bc</td>
<td>11.83 ± 0.25 b</td>
<td>8.98 ± 0.20 a</td>
<td>5.27 ± 0.13 a</td>
</tr>
<tr>
<td>BA 50</td>
<td>82.25 ± 1.59 c</td>
<td>82.33 ± 2.19 ab</td>
<td>36.44 ± 0.74 bcd</td>
<td>0.72 ± 0.01 de</td>
<td>10.68 ± 0.04 cd</td>
<td>8.23 ± 0.28 bc</td>
<td>4.97 ± 0.24 ab</td>
</tr>
<tr>
<td>BA 100</td>
<td>90.71 ± 1.71 b</td>
<td>46.00 ± 2.65 f</td>
<td>33.17 ± 1.06 d</td>
<td>0.72 ± 0.04 de</td>
<td>10.17 ± 0.10 de</td>
<td>6.73 ± 0.07 d</td>
<td>4.26 ± 0.03 cd</td>
</tr>
<tr>
<td>BA 150</td>
<td>107.50 ± 4.91 a</td>
<td>21.00 ± 2.08 g</td>
<td>27.50 ± 0.87 e</td>
<td>0.67 ± 0.03 f</td>
<td>8.50 ± 0.29 f</td>
<td>6.31 ± 0.17 d</td>
<td>3.80 ± 0.33 d</td>
</tr>
<tr>
<td>Ethrel 50</td>
<td>70.97 ± 0.21 f</td>
<td>78.67 ± 2.03 bc</td>
<td>42.14 ± 0.08 a</td>
<td>0.86 ± 0.05 a</td>
<td>12.83 ± 0.10 a</td>
<td>8.64 ± 0.04 ab</td>
<td>4.67 ± 0.20 bc</td>
</tr>
<tr>
<td>Ethrel 100</td>
<td>78.63 ± 3.25 cd</td>
<td>75.67 ± 1.86 cd</td>
<td>38.70 ± 3.52 abc</td>
<td>0.80 ± 0.01 b</td>
<td>12.88 ± 0.65 a</td>
<td>8.66 ± 0.16 ab</td>
<td>4.66 ± 0.22 bc</td>
</tr>
<tr>
<td>Ethrel 150</td>
<td>79.20 ± 1.85 ed</td>
<td>59.67 ± 2.91 e</td>
<td>37.75 ± 0.72 bc</td>
<td>0.71 ± 0.03 ef</td>
<td>9.90 ± 0.06 e</td>
<td>8.22 ± 0.02 bc</td>
<td>4.51 ± 0.48 bc</td>
</tr>
</tbody>
</table>

Note. BA: Benzyladenine; GA3: Gibberellic acid; Means sharing the same letter do not differ by DMR test at 5% probability level; ± means SE.
Table 3. Effects of growth regulators on corm and cormels associated variables of gladiolus

<table>
<thead>
<tr>
<th>Treatments (ppm)</th>
<th>Corms per plant</th>
<th>Corm diameter (cm)</th>
<th>Corm weight (g)</th>
<th>Comels per plant</th>
<th>Total cormel wt (g)</th>
<th>Av. Cormel wt (g)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>1.01 ± 0.01 d</td>
<td>5.81 ± 0.20 abc</td>
<td>57.70 ± 4.88 bc</td>
<td>8.57 ± 1.02 e</td>
<td>3.08 ± 0.75 c</td>
<td>0.41 ± 0.072 a</td>
</tr>
<tr>
<td>GA3 50</td>
<td>1.21 ± 0.04 cd</td>
<td>5.90 ± 0.05 ab</td>
<td>59.47 ± 3.13 abc</td>
<td>18.17 ± 0.29 cd</td>
<td>3.90 ± 0.08 bc</td>
<td>0.28 ± 0.071 b</td>
</tr>
<tr>
<td>GA3 100</td>
<td>1.40 ± 0.16 c</td>
<td>6.01 ± 0.05 a</td>
<td>68.30 ± 2.41 a</td>
<td>23.48 ± 2.20 b</td>
<td>4.58 ± 0.90 b</td>
<td>0.24 ± 0.032 bcd</td>
</tr>
<tr>
<td>GA3 150</td>
<td>1.25 ± 0.14 cd</td>
<td>5.59 ± 0.19 bcd</td>
<td>65.93 ± 4.77 ab</td>
<td>31.33 ± 3.95 a</td>
<td>3.77 ± 0.53 bc</td>
<td>0.13 ± 0.02 d</td>
</tr>
<tr>
<td>BA 50</td>
<td>1.19 ± 0.11 cd</td>
<td>5.46 ± 0.20 c</td>
<td>54.70 ± 3.25 c</td>
<td>17.43 ± 0.74 d</td>
<td>3.42 ± 0.21 bc</td>
<td>0.17 ± 0.01 bcd</td>
</tr>
<tr>
<td>BA 100</td>
<td>1.86 ± 0.25 b</td>
<td>4.40 ± 0.16 e</td>
<td>37.11 ± 4.92 d</td>
<td>22.69 ± 0.97 bc</td>
<td>4.52 ± 0.54 b</td>
<td>0.26 ± 0.06 b</td>
</tr>
<tr>
<td>BA 150</td>
<td>2.50 ± 0.21 a</td>
<td>3.90 ± 0.06 f</td>
<td>26.79 ± 0.82 e</td>
<td>19.36 ± 1.86 bcd</td>
<td>2.84 ± 0.17 c</td>
<td>0.20 ± 0.02 bcd</td>
</tr>
<tr>
<td>Ethrel 50</td>
<td>1.15 ± 0.03 cd</td>
<td>5.37 ± 0.16 d</td>
<td>51.48 ± 2.44 c</td>
<td>19.25 ± 1.20 bcd</td>
<td>3.67 ± 0.01 bc</td>
<td>0.14 ± 0.05 cd</td>
</tr>
<tr>
<td>Ethrel 100</td>
<td>1.14 ± 0.01 cd</td>
<td>5.69 ± 0.22 abcd</td>
<td>66.51 ± 1.51 ab</td>
<td>30.59 ± 0.01 a</td>
<td>4.17 ± 0.09 bc</td>
<td>0.23 ± 0.07 bcd</td>
</tr>
<tr>
<td>Ethrel 150</td>
<td>1.08 ± 0.05 d</td>
<td>5.91 ± 0.09 ab</td>
<td>60.10 ± 4.73 abc</td>
<td>28.79 ± 0.84 a</td>
<td>5.96 ± 0.06 a</td>
<td>0.24 ± 0.06 bc</td>
</tr>
</tbody>
</table>

Note. BA: Benzyladenine; GA3: Gibberellic acid; Means sharing the same letter do not differ by DMR test at 5% probability level; ± means SE.

Figure 2. Effect of growth regulators on multiple shoot induction and corm multiplication in gladiolus, shoot induction (A1. Control., B1,C1. 150 ppm BA treated plants), corm multiplication (A2. Control., B2,C2. 150 ppm BA treated plants)
Table 4. Pearson correlation coefficients among vegetative, floral and corm associated variables

<table>
<thead>
<tr>
<th>Variables</th>
<th>Sprouts corm</th>
<th>Veg. Plant Ht</th>
<th>Total Plant Ht</th>
<th>Days to Flower</th>
<th>Flower(%)</th>
<th>Spike length</th>
<th>Florets spike&lt;sup&gt;-1&lt;/sup&gt;</th>
<th>Floret dia</th>
<th>Floret wt</th>
<th>Corms plant&lt;sup&gt;-1&lt;/sup&gt;</th>
<th>Corm dia</th>
<th>Corm wt</th>
<th>Cormels plant&lt;sup&gt;-1&lt;/sup&gt;</th>
<th>Total corm wt</th>
</tr>
</thead>
<tbody>
<tr>
<td>Veg. Plant Ht</td>
<td>-.426*</td>
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<td>Total Plant Ht</td>
<td>-.500</td>
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<td>Days to flower</td>
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<td>.787</td>
<td>-.844</td>
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<tr>
<td>Spike length</td>
<td>-.744</td>
<td>.696</td>
<td>.800</td>
<td>-.838</td>
<td>.763</td>
<td>**</td>
<td>**</td>
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<tr>
<td>Spike diameter</td>
<td>-.459</td>
<td>.732</td>
<td>.873</td>
<td>-.729</td>
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<tr>
<td>Florets per spike</td>
<td>-.498</td>
<td>.818</td>
<td>.907</td>
<td>-.731</td>
<td>.739</td>
<td>.847</td>
<td>.868</td>
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<tr>
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<td>.672</td>
<td>.791</td>
<td>-.860</td>
<td>.821</td>
<td>.781</td>
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<td>Floret weight</td>
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<td>.452</td>
<td>-.504</td>
<td>.466</td>
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<td>-.493</td>
<td>-.618</td>
<td>.849</td>
<td>-.776</td>
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<td>-.521</td>
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<td>-.784</td>
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<tr>
<td>Corn diameter</td>
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<td>.549</td>
<td>-.821</td>
<td>.790</td>
<td>.625</td>
<td>.438</td>
<td>.526</td>
<td>.801</td>
<td>.421</td>
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<td>Corn weight</td>
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<td>Cormels per plant</td>
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<td>.255</td>
<td>-.037</td>
<td>-.038</td>
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<td>.027</td>
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<td>Total cormel weight</td>
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<td>.338</td>
<td>-.021</td>
<td>-.204</td>
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<td>.299</td>
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<td>Average Cormel wt</td>
<td>-.2</td>
<td>.133</td>
<td>-.251</td>
<td>-.088</td>
<td>.018</td>
<td>-.146</td>
<td>-.125</td>
<td>-.259</td>
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<td>-.239</td>
<td>-.143</td>
<td>.198</td>
<td>.075</td>
<td>-.449</td>
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Note. *: Correlation is significant at the 0.05 level; **: Correlation is significant at the 0.01 level; Ht: height; dia: diameter; wt: weight; Veg: vegetative.

4. Discussion

Soaking of corms in different solutions of PGRs influenced the sprouting of buds and, also increased the vegetative and total plant height. Usually only one bud sprouted from a corm of gladiolus, but application of benzyl adenine lead to sprouting of multiple sprouts from a single corm in the present research. Benzyl adenine is an important cytokinin and has the capability of multiple shoot induction in several plants and also known as branching agent. The results also confirm the findings of Carey (2008) that exogenous application of benzyladenine promotes multiple shoots in several ornamental plants, including *Petunia*, *Sempervivum*, and *Salvia*. Furthermore, Wróblewska and Dębicz (2013) also reported the increase in the number of lateral buds through the application of benzyladenine in *Portulaca umbraticola* plants.

The gibberellic acid and ethrel increased the plant height in relatively lower doses and showed reverse effect in their highest concentrations used in this study. The involvement of gibberellic acid in stem elongation process is reported by Sun and Gubler (2004) which are in line with our findings in gladiolus plants. This growth simulative effect of ethrel on plant height and corm production has also been observed by Vijay et al. (2006) in...
gladiolus and this feature has also been observed in Amsterdam, a cultivar of gladiolus, in the present study. Pre-plant soaking of corms in PGRs showed alterations in the floral characteristics of gladiolus spikes. The gibberellic acid caused the early flowering while benzyl adenine application delayed the flowering. Cytokinins have been reported for induction of early flowering in plants, including *Selenicereus* and *Hylocereus* (Khaimov & Mizrahi, 2006), *Gladiolus* (Sajjad et al., 2014) which opposes the results of the present study, but there are certain findings which support our results that cytokinin can delay flowering in some plants (Carey, 2008). The possible reason to oppose the present results is the difference in application methods, concentration and stage of growth as Sajjad et al. (2014) used a foliar application method which caused early flowering in gladiolus. As effects of plant growth regulators altered with the change in stages of plant growth on which they are applied (Bakhsh et al., 2011). Gibberellic acid increased the flowering percentage in the present study and these results strengthen by the research work of Gayakvad et al. (2014) that gibberellic acid increased the flowering in *Jatropha* plants. Ethrel at lower concentration used in this study reduced the flowering time while higher concentration caused delay in flowering. The undesired effect of high concentration of ethrel on flowering of impatiens plants is also reported by Tamari et al. (1998).

The application of benzyladeneine increased the number of corms which resulted to raise the corm production twice than the untreated plants. The positive effect of cytokinins to increase the yield of underground structures has been reported on *Allium* (Pogroszewa et al., 2007), *Muscari* (Puchalski et al., 1979), Red candyman, a cultivar of gladiolus (Aier et al., 2015) which also support our findings in Amsterdam cultivar of gladiolus. The soaking of corms in ethrel increased the production of corms compared to control plants and the results are in line with the findings of Roychowdhury (1989) that application of ethrel enhances the corm production in gladiolus, and also facilitate the bulb production in tuberose (Nagaraja et al., 1999).

Pre-plant soaking of corms in solutions of PGRs altered the interested variables in gladiolus plants. Application of benzyl adenine induced the multiple shoots and also increased the yield of corms which actually enhanced the multiplication rate of corms more than twice, compared to non treated corms. The research work also indicates that benzyladenine caused delay in flowering while gibberellic acid induced early flowering. The present research work indicated the interesting correlation among vegetative, floral and corm variables. Number of sprouted buds per corm was the most dominant trait found in this study which showed significant influence on the most of valuable traits, including the number of corms produced per plant, which encouraged the rapid multiplication of corms compared to conventional production system.

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**References**


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