Optimized HPLC -UV Method for Separation, Detection and Quantification of Endocrine Disrupting Estrogens in Low Quality Water

Sijaona Cassian Msigala¹, Faith P Mabiki¹, Bjarne Styrishave², Robinson H Mdegela³

¹Department of Physical Sciences, Faculty of Science, Sokoine University of Agriculture, P.O. Box 3038, Morogoro, Tanzania
²Department of Pharmacy, Faculty of Health and Medical Sciences, University of Copenhagen, 2100 Copenhagen OE, Denmark
³Department of Veterinary Medicine and Public Health, College of Veterinary and Medical Science, Sokoine University of Agriculture, P.O. Box 3021, Morogoro, Tanzania

Correspondence: Sijaona Cassian Msigala, Department of Physical Sciences, Faculty of Science, Sokoine University of Agriculture, P.O. Box 3038, Morogoro, Tanzania. E-mail: sijaona99@suanet.ac.tz

Received: April 5, 2017   Accepted: May 3, 2017   Online Published: May 14, 2017
doi:10.5539/ijc.v9n3p19   URL: https://doi.org/10.5539/ijc.v9n3p19

Abstract
Endocrine disrupting estrogens are emerging contaminants in aquatic ecosystems and environment in general. There are no guidelines for routine monitoring of these chemicals, despite the existing evidences of their adverse health effect to living organisms at low concentrations. This study aimed at developing and validating an optimized HPLC-UV method for detection and quantification of estradiol and ethinylestradiol. Isocratic elution was used for separation and detection of ethinylestradiol and estradiol. The mobile phase was applied with A; water B; acetonitrile (50:50) at flow rate of 0.7mL/min and injection volume 10mL. The precision and accuracy of the method were within the acceptable range. Relative standard deviation of peak area for E2 ranged from 1.373 to 3.668%, and for EE2 ranged from 0.829 to 6.495 %. The percentage recovery for E2 ranged from 82.3 to 99.84 %, and for EE2 ranged from 84.6 to 103.52 %. Linearity of the method was realized at range of 2.5 to 50 ng/mL and 100 to 1000 ng/mL for both E2 and EE2. The linear regression coefficients were 0.9979 and 0.9973 for E2 whereas for EE2 were 0.9983 and 0.9976. Limit of detection were found to be 0.05 ng/mL and 0.08 ng/mL for E2 and EE2 respectively. The obtained limits of quantification were 0.18 and 0.28 ng/mL for E2 and EE2 respectively. In untreated sewage the concentrations of E2 and EE2 were 0.28 ng/ml and 0.18 ng/ml respectively. But in subsequent wastewater stabilization ponds the concentrations were below detection limit. Therefore, the optimized HPLC-UV method is suitable for detection and quantification of endocrine disrupting estrogens when a level of pollution is at least 0.15 ng/ml. At low extent of pollution would require use of the method in conjunction with ELISA technique.

Keywords: chromatographic method, development, validated, estrogens, analytical chemistry

1. Introduction
Estrogen hormones are emerging pollutants in water. There are natural and synthetic estrogens; the natural estrogens are not only produced by humans but also by other vertebrates as well as invertebrates particularly insects (Das, 2016). The synthetic estrogens are used for birth control, hormone replacement therapies, cancer drugs and other pharmaceuticals (Shook, 2011; Sood et al., 2014). Usually the estrogens are excreted from humans and animals mainly as conjugates of glucuronide or sulphate and a lesser amount as parent compounds in urine and faeces. The conjugates undergo deconjugation to form free estrogens by enzymatic action of bacteria or fungi (Kumar et al., 2012). The occurrence of these hormones in the environment is receiving considerable attention due to the fact that most of these compounds are endocrine disrupting chemicals (Christiansen et al., 2002; Cui et al., 2006; Zheng et al., 2013; Huang et al., 2016). Several studies have demonstrated that wastewater treatment systems are the main source of estrogens to aquatic environment (Ingrand et al., 2003; Swart & Pool, 2007; Yoon et al., 2012; Belhaj et al., 2014). A reliable analytical technique is necessary for separation, detection and quantification of the estrogens in the environmental matrix such as water.

In literature there are several analytical methods for detection and quantification of estrogens from environmental samples but the methods differ in sensitivity (Ingerslev & Halling-Sørensen, 2003). Among those methods are HPLC, LC-MS, GC-MS, and immunochemical methods. The LCMS/MS and GCMS/MS are advanced methods and most
sensitive for detection of estrogens (Faqehi et al., 2016), however their use is limited by high costs. Immunochemical methods are also sensitive but have poor selectivity compared to LCMS/MS and a single assay can only detect a single steroid (Ingerslev & Halling-Sørensen 2003; Faqehi et al., 2016).

High performance liquid chromatography is a dominant analytical technique due to its applicability to diverse analytes (Dong, 2013). The technique can quantify compounds in complex samples and can be coupled with different detectors such as UV, diode array detection (DAD), evaporation light scattering detection (ELSD), refractive index detector, fluorescence detection and mass spectrometry (MS) (Dong, 2013). The successful use of HPLC requires the right combination of variety of operating conditions such as the column length and diameter, column temperature, type of the column packing, type of the mobile phase, mobile phase flow rate, and injection volume (Settle, 2004). This implies that HPLC method development and validation are inevitable for successful application of HPLC.

Method validation ensures consistent capabilities of an analytical method by defining and confirming the analytical requirement for a specific application (Magnusson & Ornemark, 2014). The ISO/IEC 17025:2005 (2005), a guideline for competence testing and laboratory calibration, stresses that method validation is necessary requirement in analytical chemistry since it demonstrate that the method is fit for the purpose. Method validation include assessment of the parameters such as precision, linearity, accuracy, limit of detection, limit of quantification, specificity, range and robustness of the method (ICH, 2005; Magnusson & Ornemark, 2014).

Schellinger & Carr (2006) found that gradient elution gave a shorter overall analysis with similar resolution compared to isocratic elution. On the other hand isocratic elution remain preferable when the sample contains less than ten weakly retained components (Schellinger & Carr, 2006).

This study aimed at establishing an optimized HPLC-UV method for separation, detection and quantification of estrogens hormones from low quality water particularly from wastewater stabilization ponds. The study focused on the two most potent estrogens namely ethinylestradiol and estradiol.

2. Materials and Methods

2.1 Chemicals and Materials

Ethinylestradiol (EE2) and β-estradiol (E2) standards hormones were supplied by Santa Cruz Biotechnology, Texas, USA. HPLC grade methanol, acetonitrile and water (99.9% purity) were supplied by Sigma Aldrich, Germany, n-heptane (99%), acetone (99.8%) and hydrochloric acid (37%, 1.18M) were supplied by Carlo Erba Reagenti. Solid phase extraction C-18 cartridges (130 mg, 3 mL) by Varian and Chromabond, silica gel (3 mL/500 mg) from Macherey-Nagel in Germany were employed.

2.2 Preparation of Standard Solution and Calibration Standard Solution

Stock solution of 10,000 ng/ml in methanol was prepared for a mixture standard E2 and EE2, and then stored at 4°C in a fridge. Serial dilution of the stock solution was carried out to obtain 1000, 750, 500, 250, 100, 50, 25, 10, 5, 2.5, 1, 0.5, and 0.1 ng/ml for the mixture of the two hormones. In addition, separate standard solutions of the two hormones were prepared at 100 ng/ml for determination of their retention time.

2.3 Spiking Standard Estrogens in Distilled Water

For determination of recoveries the estrogens standard solutions were spiked in 1 L of distilled water to make different concentrations for both E2 and EE2. The established concentrations were 1000, 750, 500, 250, 100, 50, 25, 10, 5, 2.5, 1, 0.5, and 0.1 ng/ml. Each solution was filtered twice using by GFCs, a procedure which could be applied for removal of debris from low quality water.

2.4 HPLC Conditions

Analyses were performed at Ecotoxocology and Natural Product Laboratory at the College of Veterinary and Medical Sciences, Sokoine University of Agriculture. The HPLC (Shimadzu 20AD) fitted with an auto sampler and a SPA-UV detector at 230 nm was used for analysis. A reversed phase phenomenex C-18 column (150 x 4.6 mm and particle size of 5μm) was used for separation with the oven temperature set at 35 °C. The sample injection volume was 10 μL and flow rate 0.7 mL/min. The mobile phases employed were A; water; acetonitrile (50:50).

2.5 Method Development

With the theoretical background, detection, separation and quantification of the two estrogens hormones were achieved by manipulating factors such as wavelength, mobile phase composition, flow rate and temperature. Several trials were carried out to separate, identify and quantify a mixture of estradiol, and ethinylestradiol.

When the mobile phases were; A- water and B- methanol, the hormones were detected at shorter retention time but the resolution was very poor even after several trials upon changing column temperature, flow rate and gradient. When the
mobile phases; A-water and B-acetonitrile, retention time increased with good resolution. It was observed that even when gradient elution was employed good resolution was obtained when equal proportions of the mobile phases run for longer time. Isocratic elution was found to be suitable over the gradient elution. Retention time for estradiol was 11.89 ± 0.03 minutes and for ethinylestradiol 14.79 ± 0.04 minutes while the method running time was 16 minutes.

2.6 Method Validation
Validation of method developed was carried out in accordance to ISO/IEC 17025:2005. Validation characteristics of the method such as precision, accuracy, linearity, specificity, limit of detection, quantification, and robustness were assessed. The procedures employed in assessment of validation characteristics were as stipulated in ICH (2005) document.

2.7 Collection of Water Samples from Wastewater Stabilization Ponds
Wastewater samples were collected from Mafisa WSPs owned by Morogoro Municipality in Tanzania. An untreated sewage wastewater samples were drawn using cleaned 2.5 liters glass bottle during dry season. Other samples were drawn from outlet of anaerobic and facultative ponds using cleaned 2.5 liters glass bottle. After sampling, the pH of the sampled wastewater samples was adjusted to about 3 by adding concentrated hydrochloric acid (1.18M) so as to fix the estrogens (Havens et al., 2010; Hansen et al., 2011). Then, the samples were carried in cool box packed with ice packs to the Ecotoxicology and Natural Products Research Laboratory in the College of Veterinary and Medical Sciences at Sokoine University of Agriculture. In the laboratory pretreatment and solid phase extraction of estrogens were conducted within 12 hours after sample collection.

2.8 Extraction of Estrogens from Water Samples
Extraction of estrogens from water samples was carried out according to the protocol described by (Hansen et al., 2011) with some modifications customized to our laboratory settings. Each water sample (1.5 litres) was first filtered twice using GFC filters papers to ensure removal of debris. The C-18 cartridges were conditioned with 2×3 mL heptane, 3 mL acetone, and lastly with 3 mL of distilled water. Thereafter, solid phase extraction (SPE) was performed with C-18 cartridges (Bond Elut 500 mg, 3cc reservoir, Varian Agilent Technologies, USA) facilitated by vacuum manifold. After extraction the cartridges were dried in air using vacuum manifold for about half an hour. Then, elution of the analyte was achieved by using 10 mL of heptane and acetone (65:35). Thereafter, the elute was air dried at 30 °C, followed by reconstitution using 5 mL methanol. The samples were stored at –20 °C, until analysis by HPLC-UV method.

2.9 Statistical Analysis
Microsoft excel 2007 with data analysis pack was used for statistical analysis. Descriptive statistics such as standard deviation, mean and relative standard deviation were computed.

3. Results and Discussion

3.1 Method Linearity and Range
Figure 1 and 2 demonstrate that the peak areas were directly proportional to the concentrations of E2 and EE2 respectively. Both at higher concentrations (from 100 to 1000 ng/mL) and lower concentrations (from 2.5 to 50 ng/mL) showed good linearity with linear regression coefficient being 0.9979 and 0.9973 for E2 and 0.9983 and 0.9976 for EE2. Those values of regression coefficient denote excellent linear relationship between the peak area and the concentrations of the two hormones (ICH 2005; Magnusson & Ornemark, 2014).

![Figure 1. Calibration curves for standard E2 and EE2 at high concentrations](image_url)
3.2 Method Precision

Table 1 and 2 display the method precision for detection and quantification of E2 and EE2 respectively. The relative standard deviations of peak area for both E2 and EE2 were below 2% except for the lowest concentrations which was above 2% but below 10%. Adebayo & Oduwaye (2015) stated that an effective HPLC method should have a relative standard deviation less than 10% for 6 replicates. Figure 4 revealed good precision of this method when the standard solution at 100 and 250 ng/mL were analysed.

Table 1. Method Precision for detection and Quantification of E2 at Higher and Lower concentrations

<table>
<thead>
<tr>
<th>S/N</th>
<th>(250 ng/mL)</th>
<th>100 ng/mL</th>
<th>5 ng/mL</th>
<th>0.1 ng/mL</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>RT</td>
<td>PA</td>
<td>RT</td>
<td>PA</td>
</tr>
<tr>
<td>1</td>
<td>11.766</td>
<td>930,530</td>
<td>11.860</td>
<td>435,838</td>
</tr>
<tr>
<td>2</td>
<td>11.824</td>
<td>897,259</td>
<td>11.890</td>
<td>425,610</td>
</tr>
<tr>
<td>3</td>
<td>11.874</td>
<td>919,495</td>
<td>11.903</td>
<td>421,479</td>
</tr>
<tr>
<td>4</td>
<td>11.868</td>
<td>941,589</td>
<td>11.916</td>
<td>422,706</td>
</tr>
<tr>
<td>5</td>
<td>11.844</td>
<td>891,922</td>
<td>11.908</td>
<td>422,062</td>
</tr>
<tr>
<td>6</td>
<td>11.784</td>
<td>905,404</td>
<td>11.887</td>
<td>419,813</td>
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<tr>
<td>Mean</td>
<td>11.827</td>
<td>911033.17</td>
<td>11.894</td>
<td>424584.667</td>
</tr>
<tr>
<td>Std</td>
<td>0.044</td>
<td>15164.476</td>
<td>0.020</td>
<td>5830.884</td>
</tr>
<tr>
<td>RS (%)</td>
<td>0.374</td>
<td>1.665</td>
<td>0.167</td>
<td>1.373</td>
</tr>
</tbody>
</table>

Abbreviations: RT = retention time; PA= peak area; Std = standard deviation; RS = relative standard deviation
Table 2. Method Precision for detection and Quantification of EE2 at Higher and Lower concentrations

<table>
<thead>
<tr>
<th>S/N</th>
<th>250 ng/mL RT PA</th>
<th>100 ng/mL RT PA</th>
<th>5 ng/mL RT PA</th>
<th>0.1 ng/mL RT PA</th>
</tr>
</thead>
<tbody>
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<td>2</td>
<td>14.728 923,056</td>
<td>14.878 405,267</td>
<td>14.73 18,634</td>
<td>14.854 1,887</td>
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<tr>
<td>3</td>
<td>14.768 901,806</td>
<td>14.8 403,473</td>
<td>14.816 18,694</td>
<td>14.788 1,836</td>
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<tr>
<td>6</td>
<td>14.768 924,008</td>
<td>14.774 405,635</td>
<td>14.802 18,672</td>
<td>14.803 1,890</td>
</tr>
</tbody>
</table>


Std 0.046 15,090.705 0.022 3352.977 0.043 310.252 0.030 126.822

RS(%) 0.310 1.659 0.151 0.829 0.289 1.658 0.201 6.495

Abbreviations: RT = retention time; PA = peak area; Std = standard deviation; RS = relative standard deviation

3.3 Method Accuracy

Table 3 and 4 display the method performance accuracy for quantification of E2 and EE2 respectively. For E2 recoveries ranged from 82.3 to 99.84 %, on the other hand recoveries for EE2 ranged from 84.6 to 103.52 %. The recoveries are within the recommended range for analytical method between 80 % to 120 % (Shabir, 2004).

Table 3. Method performance accuracy for quantification of E2 at Low & High Concentrations

<table>
<thead>
<tr>
<th>Spiked Concentration ng/mL</th>
<th>Sample No.</th>
<th>Calculated amount ng/mL</th>
<th>% Recovery</th>
<th>Spiked Concentration ng/mL</th>
<th>Sample No.</th>
<th>Calculated amount ng/mL</th>
<th>% Recovery</th>
</tr>
</thead>
<tbody>
<tr>
<td>50</td>
<td>1</td>
<td>49.61</td>
<td>99.22</td>
<td>1000</td>
<td>1</td>
<td>954.93</td>
<td>95.49</td>
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<tr>
<td></td>
<td>2</td>
<td>49.90</td>
<td>99.80</td>
<td></td>
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<td>987.00</td>
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<tr>
<td>25</td>
<td>1</td>
<td>24.74</td>
<td>98.96</td>
<td>750</td>
<td>1</td>
<td>748.83</td>
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<td></td>
<td>2</td>
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<td>99.52</td>
<td></td>
<td>2</td>
<td>747.50</td>
<td>99.67</td>
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<tr>
<td>10</td>
<td>1</td>
<td>8.23</td>
<td>82.30</td>
<td>500</td>
<td>1</td>
<td>468.39</td>
<td>93.68</td>
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<tr>
<td></td>
<td>2</td>
<td>8.97</td>
<td>89.70</td>
<td></td>
<td>2</td>
<td>480.00</td>
<td>96.00</td>
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<tr>
<td>5</td>
<td>1</td>
<td>4.93</td>
<td>98.60</td>
<td>250</td>
<td>1</td>
<td>239.62</td>
<td>95.85</td>
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<td>2</td>
<td>5.12</td>
<td>102.40</td>
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<td>245.98</td>
<td>98.39</td>
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<tr>
<td>2.5</td>
<td>1</td>
<td>2.44</td>
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<td>2.49</td>
<td>99.60</td>
<td></td>
<td>2</td>
<td>99.45</td>
<td>99.45</td>
</tr>
</tbody>
</table>

Table 4. Method performance accuracy for quantification of EE2 at Low & High Concentrations

<table>
<thead>
<tr>
<th>Spiked Concentration ng/mL</th>
<th>Sample No.</th>
<th>Calculated amount ng/mL</th>
<th>% Recovery</th>
<th>Spiked Concentration ng/mL</th>
<th>Sample No.</th>
<th>Calculated amount ng/mL</th>
<th>% Recovery</th>
</tr>
</thead>
<tbody>
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<td>50</td>
<td>1</td>
<td>49.700</td>
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<td>98.96</td>
<td></td>
<td>2</td>
<td>1002.00</td>
<td>100.2</td>
</tr>
<tr>
<td>25</td>
<td>1</td>
<td>24.057</td>
<td>96.23</td>
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<td>753</td>
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<td>96.62</td>
<td></td>
<td>2</td>
<td>749.45</td>
<td>99.93</td>
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<tr>
<td>10</td>
<td>1</td>
<td>8.460</td>
<td>84.6</td>
<td>500</td>
<td>1</td>
<td>468.35</td>
<td>93.67</td>
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<tr>
<td></td>
<td>2</td>
<td>8.860</td>
<td>88.6</td>
<td></td>
<td>2</td>
<td>485.10</td>
<td>97.02</td>
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<td>1</td>
<td>5.176</td>
<td>103.52</td>
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<td>99.12</td>
<td></td>
<td>2</td>
<td>99.98</td>
<td>99.98</td>
</tr>
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</table>

3.4 Method Sensitivity

Limit of detection (LOD) and limit of quantification (LOQ) were determined using equation 1 and 2 respectively. The LOD and LOQ for E2 were found to be 0.054 ng/mL and 0.18 ng/mL respectively. The corresponding LOD and LOQ for EE2 were 0.084 ng/mL and 0.28 ng/mL respectively. This result implies that the method is suitable for moderate and higher extent of pollution of low quality water by estrogens. At low extent of pollution the method could be used in conjunction with ELISA technique. Huang & Sedlak (2001) used HPLC-UV method in conjunction with ELISA.
technique for analysis of estrogens in wastewater; the results were comparable to those obtained by applying GC-MS/MS. In addition, the developed HPLC-UV method can be used in low extent pollution when standard addition technique would be applied in quantifying the estrogens. The sensitivity of this method could be comparable to HPLC-DAD method which allowed precise and accurate determination of estrogens in various environmental water at concentrations ranging between 0.025 ng/ml to 0.3 ng/ml, (Lopez and Alpha, 2001). On the other hand, this method demonstrated higher sensitivity than HPLC-UV method developed by Fonseca et al., 2013, in such method the detection limits were 0.25 ng/ml and 0.11 ng/ml for E2 and EE2 respectively. Contrary, LC-MS/MS method is more sensitive than this method, for instance Naldi et al., 2016 developed LC-MS/MS method for determination of free and conjugated steroids that had LOD ranging between 0.003 ng/mL to 0.015 ng/mL for surface water and 0.014 ng/mL to 0.076 ng/mL in wastewater samples.

\[
LOD = \frac{\text{standard deviation}}{\text{slope}} \times 3 \quad \text{....equation 1}
\]

\[
LOQ = \frac{\text{Standard deviation}}{\text{slope}} \times 10 \quad \text{....equation 2}
\]

3.5 Robustness

The method gave comparable results at different wavelengths such as at 215, 230, and 254 nm. Likewise when column temperature was varied from 35°C to 40°C no significant change of result occurred. Contrary to that retention time changed with change in flow rate for instance at flow rate 1 mL/min retention time was found to be 8.3 and 10.4 minutes for E2 and EE2 respectively. Whereas at a flow rate of 0.7 mL/min the retention time is 11.89 ± 0.03 and 14.79 ± 0.04 minutes for E2 and EE2 respectively.

![Chromatogram of E2 and EE2 when concentration was 10 ng/mL](image)

Figure 3. Chromatogram of E2 and EE2 when concentration was 10 ng/mL.
3.6 Levels of Estrogens in LQW Detected through HPLC-UV Method

Estrogens were detected in untreated sewage sample but in the outlets of anaerobic and facultative ponds were below the detection limit. The concentration of E2 and EE2 in untreated sewage was found to be 0.28 ng/ml and 0.18 ng/ml respectively. When ELISA competitive technique was employed revealed that the levels of estrogens in the study area were below the detection limit of the developed method. The concentrations EE2 and E2 at Mafisa WSPs ranged from 0.036 ng/ml to 0.0015 ng/ml and 0.086 ng/ml to 0.0044 ng/ml respectively (Msigala et al., 2017).

4. Conclusion

This paper presents a fast, linear, precise, accurate and robust HPLC-UV method for separation, detection and quantification of estrogens hormones from low quality water. The method can give precise and accurate results for moderate and higher extent of pollution of low quality water by estrogens. At lower extent of pollution (below 0.15 ng/mL) can be used in conjunction with ELISA technique, in such combination the HPLC method offset the shortcomings of the results which could be obtained through ELISA alone, on the other hand utilize the high sensitivity of ELISA technique.

Conflict of Interest

The authors declare that there are no conflicts of interest which could potentially influence this work

Acknowledgments

This research was funded by Danish Ministry of Foreign Affairs through Safe Water for Food (SaWaFo) project. We are very grateful to DANIDA for funding this research.

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