Analysis of the Essential Oils of Leaves and Stems of *Crassocephalum crepidioides* Growing in South Western Nigeria

Owokotomo, I. A. (Corresponding author) Department of Chemistry, Federal University of Technology PMB 704, Akure, Nigeria Tel: 234-803-542-6559 E-mail: kunleowokotomo@yahoo.com

Ekundayo, O.

Department of chemistry, University of Ibadan, Ibadan, Nigeria Tel: 234-803-064-3828 E-mail: o.ekundayo@yahoo.com

Oladosu, I. A.

Department of chemistry, University of Ibadan, Ibadan, Nigeria Tel: 234-803-656-0184 E-mail: ai.oladosu@mail.ui.edu.ng

Aboaba, S. A.

Department of chemistry, University of Ibadan, Ibadan, Nigeria Tel: 234-803-801-1394 E-mail: sa.aboaba@mail.ui.edu.ng

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Abstract

The essential oils from fresh leaves and stems of *Crassocephalum crepidioides* (Asteraceae) growing wild in south western Nigeria and used as food supplement by the local population were extracted. The essential oils were extracted using Clevenger- type hydrodistiller and analysed by GC- MS. The percentage yields of oils were 0.12 %v/w and 0.09 %v/w, respectively. The essential oils of the leaves contained α -caryophyllene (10.29 %), β -cubebene (13.77 %) and α -farnesene (13.27 %) as major constituents while the dominant constituents of the stems oil were thymol (43.93 %), α -caryophyllene (15.16 %) and 4-cyclohexybutyramide (20.94 %).

Keywords: Essential oil, Crassocephalum crepidioides, Hydrodistillation, South western Nigeria

1. Introduction

Crassocephalum crepidioides (Asteraceae) known locally in south western Nigeria as "Ebolo" is a popular food supplement in Nigeria (Agbogidi, 2010). The plant grows in large quantity as undercover in tree crop plantations (Dairo et al., 2007). The edible leaves and stems are used to treat indigestion or as laxative, as purgative and remedy for lever problem (Fowomola et al., 2005; Ayodele, 2007). The antioxidant activity of *Crassocephalum crepidioides* has been demonstrated. The plant extracts showed marked antioxidant activity in linoleic acid model systems (Odukoya et al., 2007).

Free radical scavenging and Hepatoprotective actions of *Crassocephalum crepidioides* has also been demonstrated in Japan where the plant grows wildly in the Okinawa Islands and well known among folks as remedy for acute hepatitis and fever (Yoko Aniya et al., 2005). Also, extracts from the plant have been shown to possess cancer chemo preventive actions and anti-inflammatory properties (Chia- Chung Hou et al., 2007).

The chemical constituent of the ethanol extracts and the lipid peroxidation inhibitory effect of *C. Crepidiodes* from Nigeria has been investigated (Salawu et al., 2006). The result reveals the presence of phenyl acetic acid,

Vanillic acid, protocatechiuc acid and caffeic acid. The plant extracts also showed high tendency to inhibit lipid peroxide formation and thus capable of preventing its numerous pathological effects.

Literature reports on the essential oils composition of *C. cepidioides* are scant. However, Zollo et al. (2000) reported that the essential oils of *C. crepidiodes* and six other species of the same genus from Cameroon were composed mainly of α -phellandrene, p-cymene, pinene, myrecene, limonene and E- β -ocimene as the major constituents.

Essential oils of a large number of plants possess useful biological, pharmacological and therapeutically activities and are commercially important compounds. Their utilization in the various industries is influenced by the nature of their constituents (Ekundayo, 1986).

It is known that the essential oils composition pattern could be affected by the geographical and climatic conditions(Arras et al., 2009), and to the best of our knowledge there has been no previous report on the chemical compositions of essential oils of *C. crepidioides* from Nigeria. On these bases, we investigate the essential oils of leaves and stems of *Crassocephalum crepidioides* indigenous in south western Nigeria.

2. Materials and Methods

2.1 Plant material and Isolation of Essential Oils

The plant materials used for this experiment were harvested at an abandoned cocoa plantation behind The Federal University of Technology, Akure, Ondo state, Nigeria. The identification was done by experts at the Forest research institute of Nigeria (FRIN) where Voucher specimen was deposited in the Herbarium. Fresh plant parts were carefully separated, washed and then subjected to hydro- distillation separately for 1-2hrs using a Clevenger-type glass apparatus, according to British Pharmacopoeia (1980) specification, but modified with graduated column to enable direct estimation of oil yield. The weight of fresh plant material was 1 Kg for each plant part. The oil yield was calculated in percentage of volume per weight of plant samples. The oil samples were stored in air-tight containers at 0 $^{\circ}$ C before going for GC-MS analysis without any further treatment.

2.2 Gas Chromatography/mass Spectrometry Analysis

The essential oils were analysed using GC-MS Agilent 6890N GC Coupled with MS-5973-634071 Series. The capillary column type was DB-1MS [30.0 m (length) X 320.00 μ m (diameter) X 1.00 μ m (film thickness)]. The carrier gas was Helium at constant flow rate of 1.0 ml/min and average velocity of 37 cm/s; the pressure was 0.78 psi. The initial column temperature was set at 100 °C (hold for 2 min) to the final temperature of 250 °C at the rate of 5 °C /min, Volume injected was 1.0 μ L and split ratio was 50:1.

The total chromatogram was auto-integrated by ShemStation and the constituents were identified by comparison with published mass spectral database (NIST02.L) and data from literature.

3. Results and Discussion

3.1 Percentage Yield

The percentage yield of essential oil of leaves and stems of C. crepidioides were 0.12 % and 0.09 % v/w respectively.

3.2 Chemical Constituents

A total of thirty-five compounds were identified in the essential oils of the leaves and stems of *C. crepidioide*. The stems essential oil yielded just five compounds against thirty-two compounds obtained from the leaves essential oils. Thymol (43.93 %), 4-cyclohexyl butyramide (20.94 %) and α -caryophyllene (15.16 %), α + β -caryophyllene (10.72 %) and 6-Phenyl dodecane (9.25 %) were the compounds present in the stems oil, while β -cubebene (13.77 %), α -farnesene (13.27 %) and α -caryophyllene (10.29 %) form the major constituents of the leaves essential oil. Other notable constituents of the leaves essential oil were Phytol (6.30 %), caryophyllene (6.05 %), Z-1-(1-butenyl) aziridine (3.73 %) and cis- β -farnesene (3.06 %). α -caryophyllene was the only constituent common to both the stems and the leaves essential oils of *C. crepidiodes*. Thus the leaves serve as the major reservoir of chemical compounds in the plant. Complete list of compounds obtained from the stems and leaves of *C. crepidioides* are summarised in Table 1.

4. Conclusions

In conclusion, the essential oils of *Crassocephallum crepidiodes* indigenous in south western Nigeria afforded a plethora of compounds of which thymol, α -caryophyllene, β -cubebene, α -Farnesene and 4-cyclohexybutyramide were the major constituents. The study reveals that the composition of the oils differ from the earlier report from

Cameroon and may therefore be regarded as a different chemotype. *C. crepidiodes* growing in South Western Nigeria may be utilised as a natural source for thymol which is known to possess antimicrobial activity.

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Chemical compounds	Retention	Percentage composition (%)	
	Times (Min)	Leaves	Stems
m-tert-Butyl phenol	2.25	2.44	-
P-Myrecene	2.41	0.80	-
1-Methyl-2-(1-methylethyl)- 3-(1-methylethylidene)cyclopropane	2.47	1.08	-
Myrtenol	2.53	2.78	-
2,6-Dimethyl-3,5,7-octatriene-2-ol	3.09	2.45	-
2-lsopropylidene-3-methylhexa-3,5dienal	3.67	1.06	-
Z-1-[1-butenyl]aziridine	3.80	3.73	-
Thymol	4.08	-	43.93
3-Methylene-p-menth-8-ene	5.37	1.12	-
Artemisia triene	5.56	2.78	-
Copaene	5.70	1.71	-
2-Dodecanone	5.80	2.78	-
β-Elemene	5.94	2.24	-
Caryophyllene	6.40	6.05	-
α-Caryophyllene	7.00	10.29	15.16
α+β-Caryophyllene	7.12	-	10.72
cis-β-Farnesene	7.14	3.06	-
β-Cubebene	7.51	13.77	-
2-Tridecanone	7.73	2.53	-
β-Elemene	7.93	3.17	-
α-Fernesene	8.15	3.27	-
δ-Cadinene	8.32	1.35	-
γ-Elemene	8.88	1.83	-
3,7,11-Trimethyl-1,6,10-dodecatrien-3-ol	9.11	2.89	-
Caryophyllene oxide	9.23	2.18	-
Humulene epoxide II	9.17	1.07	-
α-Santalol	10.15	0.71	-
Z-Ocimene	10.32	2.35	-
2-Methylene-6,8,8-trimethyl tricycle[5.2.2.0(1,6)]undecan-3-ol	11.21	1.07	-
2,6,8-Trimethyl decanoic acid methylester	13.65	0.73	-
6-Phenyl dodecane	14.50	_	9.25
1,19-Eicosadione	14.99	0.85	-
Phytol	16.50	6.30	-
n-Hexadecanoic acid	17.14	1.06	-
4-Cyclohexylbutyramide	23.28	20.94	-

Table1. Chemical composition of the essential oil of leaves and stems of Crassocephallum crepidioides