

Genetic Perturbation of Redox Status and Fructose-Supported Elevation of Dark Anaerobic Hydrogen Evolution in *Synechocystis* sp. Strain PCC 6803

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Abstract

The effects of fructose on the dark anaerobic hydrogen production of the genetically modified mutants from glucose tolerant derivative of unicellular cyanobacterium *Synechocystis* sp. strain PCC 6803 (GT strain) in nitrate-free buffer solution has been studied by following the time courses of the amounts of hydrogen, fructose, dry cell weight, lactate and acetate per culture volume and the amount of endogenous glucose per dry cell weight. Considering that hydrogen production on bidirectional NiFe-hydrogenase competes NAD(P)H with lactate dehydrogenase and alcohol dehydrogenase, mutants which lack lactate dehydrogenase (Δ ldh mutant) and both lactate dehydrogenase and alcohol dehydrogenase (Δ adh Δ ldh mutant), were utilized. These genetic modifications have a high potential to redirect electrons to NiFe-hydrogenase for hydrogen generation. A supply of additive to increase electron supply has found to be a successful method to activate hydrogen production. Fructose was selected as an additive due to our previous observation that hydrogen production in GT strain was highly increased in the presence of 50 μ mol mL⁻¹ fructose. Initial hydrogen production rate of Δ adh Δ ldh mutant in the presence of 50 μ mol mL⁻¹ fructose was increased to 0.129 μ mol mL⁻¹h⁻¹ that was 5.11 times that of GT strain. This increase was associated with the initial increase in glucose accumulation rate. Hydrogen production by Δ adh Δ ldh mutant in the presence of 50 μ mol mL⁻¹ fructose was quite stable for at least 120 h. Fructose-mediated hydrogen production resulted in an 11-fold increase in the moles of hydrogen per culture volume at 120 h over controls.

Keywords: fructose, hydrogen, lactate dehydrogenase, alcohol dehydrogenase, NiFe-hydrogenase, nitrate-free, *Synechocystis* sp. strain PCC6803

1. Introduction

Establishment of sustainable and low emission fuels is an urgent issue due to the inevitable deprivation of petroleum resource and rising carbon dioxide emission. Cyanobacteria have attracted much attention as a biotechnical tool to convert carbon dioxide to clean fuel hydrogen because of the autotrophic growth to accumulate endogenous glucose with high growth rate, readily available genetic tools, low nutrient requirement, easiness to extract intracellular components and non-food competitor.

Synechocystis sp. strain PCC 6803 is the most studied unicellular and N₂-non-fixing cyanobacterium with complete genetic database sequence (Kaneko et al., 1996). This strain is able to produce hydrogen on bidirectional NiFe-hydrogenase under dark anaerobic condition (Howarth & Codd, 1985). The enzyme is annotated as NAD(P)H dependent in several works (Cournac et al., 2004; Germer et al., 2009) and ferredoxin/ flavodoxin dependent in a recent work (Gutekunst et al., 2014). Dark anaerobic and nitrate-free metabolism generates NAD(P)H from the catabolism of high energy cellular components that are stored during photosynthesis. Catabolism of stored components is realized to maintain cell viability. This destructive metabolism results in the breakdown of complex biomaterials, the release of ATP and the evolution of many metabolites such as hydrogen, lactate and acetate.

Redox homeostasis is important for living systems. Reduction and oxidation in global cellular metabolisms are coupled to buffer the toxic effect of redox unbalance by gaining and losing electrons. Cellular energy metabolism is directed by NADH and NAD⁺, and biosynthetic pathways and antioxidant defense are regulated by their phosphorylated forms NADPH and NADP⁺. Redox homeostasis is regulated at the subcellular level, hence analysis of it in multicellular organism level presents a great deal of difficulties. Despite this fact, some studies

have recently challenged the study on redox homeostasis utilizing computational analysis (Elvitigala et al., 2007) and experiments focusing on reactive oxygen species (ROS) and glutathione with unicellular cyanobacterium *Synechocystis* sp. strain PCC6803 (Guo et al., 2014; Cameron & Pakrasi, 2010). Cyanobacterium is a photolithotroph that requires and captures radiant energy on thylakoid membrane utilizing water as electron donor and produces oxygen molecules, protons, electrons and subsequently NADPH and ATP. The generation of oxygen is crucial for discharging accumulated electrons. Above works have dealt with the harmful reactive oxygen species (ROS) The accumulated NADPH, protons and electrons are utilized in the Calvin cycle to buffer redox unbalance. Previous redox homeostasis studies on cyanobacteria have concerned with the biological status under the aerobic radiated condition, and there are remarkably few data available on redox homeostasis of cyanobacteria under anaerobic dark condition. Expression of NiFe-hydrogenase gene in *Synechocystis* sp. strain PCC 6803 has been proven to be essential for dark anaerobic mixotrophic growth on glucose (Gutekunst et al., 2014).

In our laboratory work, we have pursued several research works to characterize hydrogen production in *Synechocystis* sp. strain PCC6803. The amount of hydrogen produced from *Synechocystis* sp. strain PCC6803 is varied by how the cells have been prepared in photosynthesis prior to dark anaerobic fermentation. Compositions of ions for cell growth in photosynthesis highly affect hydrogen production in dark anaerobic condition (Yamamoto et al., 2012a). Dark anaerobic hydrogen production of GT strain was activated in the presence of some monosaccharide (Chongsuksantikul et al., 2014c). Cells of GT strain prepared in the photosynthesis medium with glucose and those dark incubated in the buffer solution with glucose yielded the higher amount of hydrogen per culture volume at the end of incubation (Yamamoto et al., 2012b). Glucose was utilized for cell growth. Among several monosaccharides, fructose has proven to greatly promote hydrogen production in GT strain. The limited heterotrophic growth of GT strain on fructose in buffer solution was observed (Chongsuksantikul et al., 2014b). To increase the supply of reductive compounds NAD(P)H, the mutant lacking D-lactate dehydrogenase (Δ addh mutant) and that lacking both D-lactate dehydrogenase and alcohol dehydrogenase (Δ adh Δ addh mutant) were constructed (Yamamoto et al., 2012c; Chongsuksantikul et al., 2014a). The initial rates of dark anaerobic hydrogen production in genetically modified mutants were increased from GT strain, while the amount of hydrogen per culture volume at 96 h of Δ addh mutant was comparable with that of GT strain and that of Δ adh Δ addh mutant was lower than that of GT strain and Δ addh mutant. Redox homeostasis network appears to be changed by genetic modification of NAD(P)H related reactions. Those mutants represent a high hydrogen production potential, however, for increasing attainable level of the amount of hydrogen per culture volume, it is unavoidable to solve the problem of deactivation of NiFe-hydrogenase in mutants during dark incubation.

Thus the primary objective of this study was to stabilize long-term hydrogen production in genetically modified Δ addh mutant and Δ adh Δ addh mutant of glucose tolerant derivative of *Synechocystis* sp. strain PCC6803 utilizing fructose.

1.1 Preliminary Remarks

The glucose tolerant mutant of *Synechocystis* sp. strain PCC6803 generates molecular hydrogen by NiFe-hydrogenase, when cells aerobically and photosynthetically grown on BG-11 medium are incubated. The hydrogen production activity of anaerobic dark nitrate-free buffer solution is defined by

$$a = \frac{r_{H_2}}{r_{H_2,0}} \quad (1)$$

in which r_{H_2} ($=dy_{H_2}/dt$) and $r_{H_2,0}$ are rate of hydrogen production at time t and that with a fresh inoculum. If first-order deactivation is assumed, the kinetic expression of Eq.(1) becomes

$$r_{H_2} = \frac{dy_{H_2}}{dt} = r_{H_2,0} a = r_{H_2,0} \exp(-kt) \quad (2)$$

in which k is the deactivation constant. Integration of Eq.(2) results that

$$y_{H_2} = \frac{r_{H_2,0}}{k} \{1 - \exp(-kt)\} \quad (3)$$

Previous studies have shown that the y_{H_2} estimated by Eq.(3) fits well with observed y_{H_2} (Chongsuksantikul et al. 2014c). Table 1 tabulates the model parameters $r_{H_2,0}$ and k that were reported in our previous works. Figure 1 shows the simulated time courses of y_{H_2} in GT strain (solid line), Δ addh mutant (broken line) and Δ adh Δ addh mutant (long dashed short dashed line) incubated in HEPES buffer solutions and in GT strain (solid line) incubated in HEPES buffer solution with 28 $\mu\text{mol mL}^{-1}$ fructose. Initial hydrogen production rate of Δ addh mutant or Δ adh Δ addh

mutant is higher than that of GT strain, while the attainable level of y_{H_2} ($y_{H_2,f}$) of $\Delta addh$ mutant is comparable with that of GT strain and that of $\Delta adh\Delta addh$ mutant is lower than that of GT strain or $\Delta addh$ mutant.

Table 1. Model parameters in Eq.(3) shown in previous reports of dark anaerobic hydrogen production by cells of GT strain, $\Delta addh$ mutant and $\Delta adh\Delta addh$ mutant of *Synechocystis* sp. strain PCC6803 in HEPES buffer solution without or with fructose. Cells were harvested at the late-logarithmic growth phase before dark anaerobic fermentation

Strain	Growth phase of inoculum	Solution	X_0 [mg mL ⁻¹]	$r_{H_2,0}$ [$\mu\text{mol mL}^{-1}\text{h}^{-1}$]	$y_{H_2,f}$ [$\mu\text{mol mL}^{-1}$]	K [h ⁻¹]	Ref.
GT strain	Late-logarithmic	H	2.1	0.0309	2.18	0.0187	Ref. 1
		H+F	2.1	0.0467	3.05	0.0221	Ref. 1
$\Delta addh$ mutant	Late-logarithmic	H	2.1	0.0359	2.31	0.0213	Ref. 2
$\Delta adh\Delta addh$ mutant	Late-logarithmic		2.1	0.0355	1.48	0.0365	Ref. 2

$\Delta addh$ mutant, a derivative of GT strain that lacks lactate dehydrogenase activity; $\Delta adh\Delta addh$ mutant, a derivative of GT strain that lacks both lactate dehydrogenase activity and alcohol dehydrogenase activity; H, HEPES buffer solution; H+F, HEPES buffer solution with 28 $\mu\text{mol mL}^{-1}$ fructose; Ref1, Chongsuksantikul et al. 2014c; Ref2, Chongsuksantikul et al. 2014a;

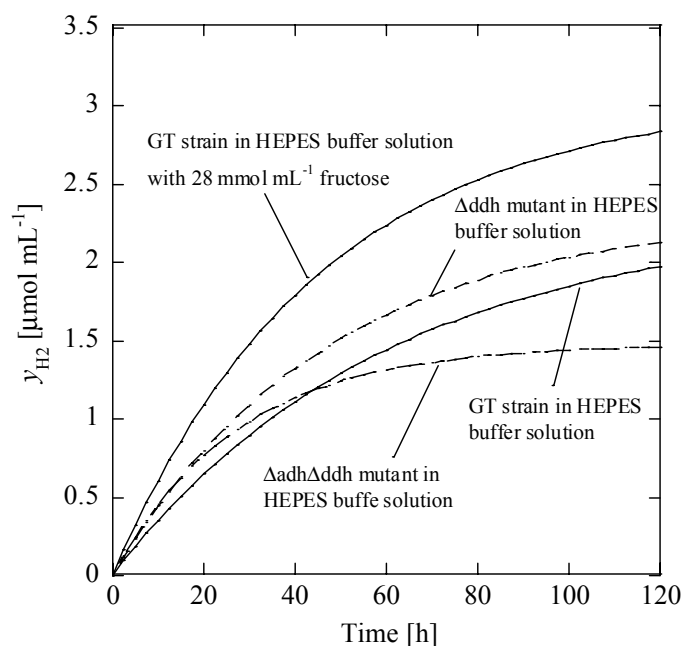


Figure 1. Estimated time courses of the number of moles of hydrogen per culture volume (y_{H_2}) for the dark anaerobic hydrogen production in GT strain (—), $\Delta addh$ mutant (---) and $\Delta adh\Delta addh$ mutant (----) utilizing Eq.(3) and parameters in Table 1. Curves for GT strain are calculated utilizing parameters for runs with HEPES buffer solution without or with 28 $\mu\text{mol mL}^{-1}$ fructose. Curves for $\Delta addh$ mutant and $\Delta adh\Delta addh$ mutant are calculated utilizing parameters only for runs with HEPES buffer solution

2. Materials and Methods

2.1 Strains

The glucose tolerant mutant of *Synechocystis* sp. strain PCC6803 (GT strain) was kindly supplied by Prof. Hihara. The mutant of GT strain that lacks D-lactate dehydrogenase activity ($\Delta addh$ mutant) and both

D-lactate dehydrogenase and alcohol dehydrogenase activities ($\Delta adh\Delta addh$ mutant) were previously constructed (Chongsuksantikul et al. 2014a).

2.2 Photoautotrophic Growth for Inoculum Preparation

Cells of GT strain, Δ addh mutant and Δ adh Δ addh mutant of *Synechocystis* sp. strain PCC6803 were grown photoautotrophically on antibiotic-free BG-11 medium (initial pH 7.8) that was aerated by 6% carbon dioxide in air in clear Pyrex glass test tube (TE-32, AGC TECHNO GLASS Co. Ltd.) at 34 °C in water bath, surface of which was exposed to fluorescent lamps with a photosynthetic photon flux density (PPFD) of 100 $\mu\text{mol-photons m}^{-2} \text{s}^{-1}$. Growth was monitored by measuring OD₇₃₀ of cell suspension. One unit of OD₇₃₀ represents the dry cell weight concentration (X) of 0.369 mg mL⁻¹. In preliminary experiments it was observed that cells were in logarithmic growth phase when OD₇₃₀ of cell suspensions was 3. Cultures of OD₇₃₀ at 3 were centrifuged at 25 °C, 3000 rpm for 10 min.

2.3 Dark Anaerobic Incubation for Hydrogen Production

The cell pellets were washed, re-suspended and incubated in 10 mL of 50 mmol L⁻¹ HEPES (4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid) buffer (pH 7.8) in 32 mL glass test tube with butyl rubber cap. HEPES was selected for pH control preventing a drastic drop in pH during fermentation. The initial dry cell mass weight concentration was set at 2 mg mL⁻¹ with 50 $\mu\text{mol mL}^{-1}$ fructose. Cell suspension was purged with nitrogen gas for a few minutes to remove oxygen gas. Incubation of cell suspension in test tube was carried out under dark anaerobic conditions, shaking at 145 rpm in a reciprocating shaker. The reciprocating distance was 40 mm and the horizontal angle was about 30 °C. All experiments were carried out in duplicate.

The number of moles of hydrogen per culture volume (y_{H_2}) was determined by analyzing the composition of gas phase in 32 mL test tube utilizing gas chromatograph and thermal conductivity detector (GC-320, GL science Inc.; column, Molecular sieve 13X; carrier gas, nitrogen gas; column temperature, 37 °C; injector temperature, 45 °C; detector temperature 80 °C).

The amount of cellular glycogen was estimated in terms of the number of moles of glucose equivalent. The number of moles of endogenous glucose per dry cell weight (m_G) was determined as follows: the amount of dry cell weight per culture volume was determined according to previous report (Yamamoto et al. 2012); cell pellets were collected by centrifugation at 25 °C, 3000 rpm for 10 min then washed 3 times by deionized water to eliminate extracellular carbon sources; intracellular glycogen was extracted and decomposed to glucose in 50 μL of 6 N HCl at 80 °C for 30 min; glucose amount was determined by using Glucose CII Test Wako (Wako Pure Chemical Ind., Ltd.).

Extracellular concentrations of lactate (c_L), acetate (c_A) and ethanol (c_E) in a 500 μL portion of supernatants were analyzed by HPLC (JASCO PU-2080 Plus and UV-2075 Plus). Mobile phase was 18 mmol L⁻¹ KH₂PO₄, pH of which was adjusted at 2.3 with H₃PO₄. Flow rate of mobile phase was 0.7 mL min⁻¹. Column temperature was 30 °C. A 10 μL of supernatant sample was injected by using 10 μL syringes. The wave length of 210 nm was used for detection of metabolites.

Unreacted fructose concentration was analyzed by HPLC (Shimadzu LC-10AD and RID-6A) with a 6.0 \times 150 mm column SZ5532 (Showa Denko Co.). Column temperature was set at 50°C. Mobile phase was 70% acetonitrile with water. Mobile phase flow rate was set to be 0.7 mL/min. A 10 μL of supernatant sample was injected by using a 10 μL syringe.

3. Results and Discussions

Our previous works show that fructose largely elevates the dark anaerobic hydrogen production of GT strain cells from logarithmic growth phase and optimum fructose concentration regarding hydrogen production is 50 $\mu\text{mol mL}^{-1}$ (Chongsuksantikul et al., 2014b), and that, in the absence of fructose, the initial hydrogen production rate of Δ addh mutant prepared from late-logarithmic growth phase in photosynthesis is higher than that of GT strain. Dark anaerobic incubation of those cells in the presence of glucose also shows that the rate of hydrogen production by Δ addh mutant is higher than that of GT strain (Yamamoto et al., 2012c; Chongsuksantikul et al., 2014a). Therefore, a series of dark anaerobic hydrogen production experiments were performed in which cells of Δ addh mutant and Δ adh Δ addh mutant from logarithmic growth phase were incubated in HEPES buffer solution without or with 50 $\mu\text{mol mL}^{-1}$ fructose.

In Figure 2, effects of fructose upon dark anaerobic hydrogen production in GT strain, Δ addh mutant and Δ adh Δ addh mutant are shown. Cells were incubated in HEPES buffer solution or HEPES buffer solution with 50 $\mu\text{mol mL}^{-1}$ fructose. The amount of hydrogen produced from unit volume of solution (y_{H_2}), the amount of endogenous glucose in glycogen of a unit dry cell weight (m_G), dry cell weight per solution volume (X), the concentrations of fructose, lactate and acetate were determined.

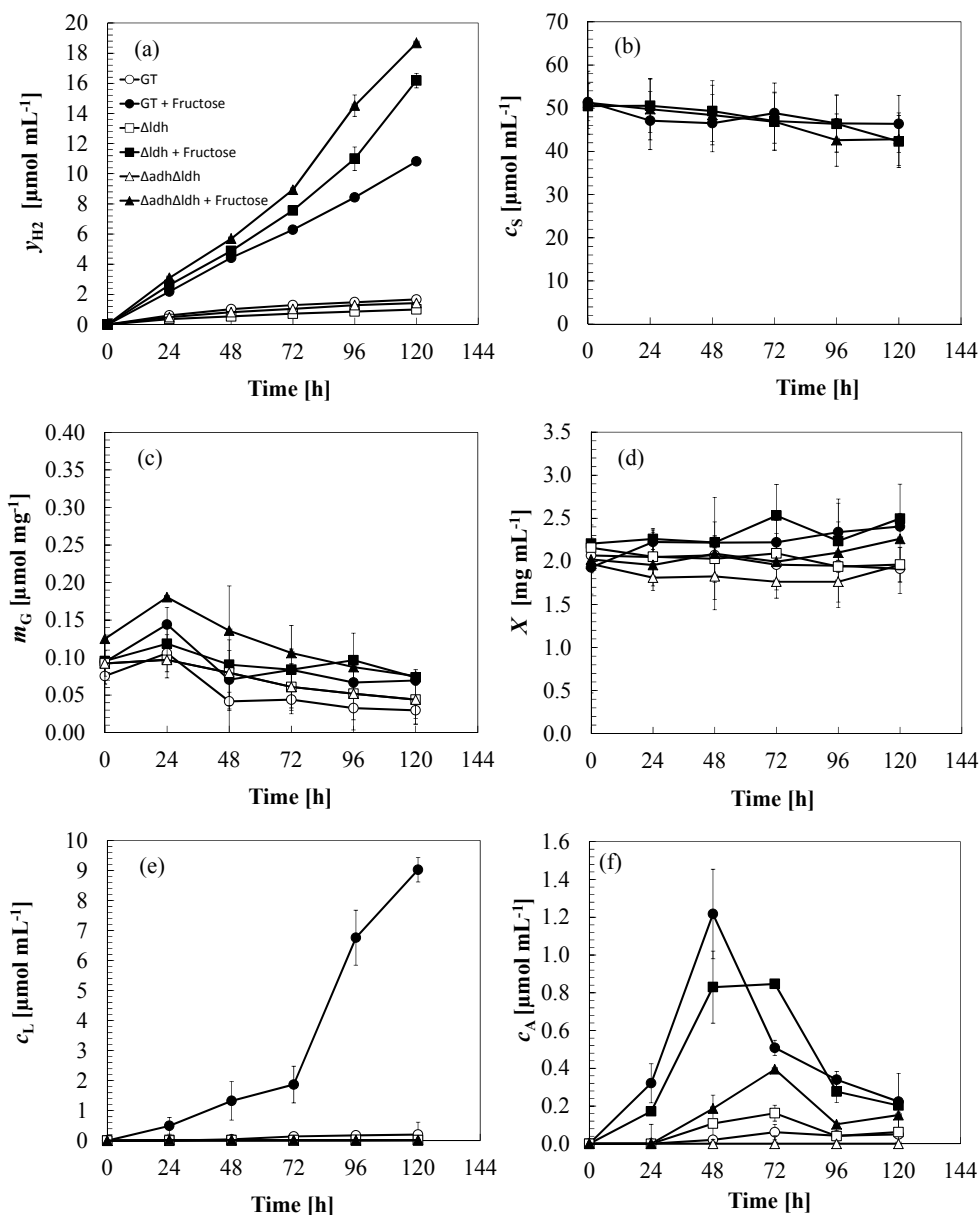


Figure 2. Time courses of culture parameters for dark anaerobic hydrogen production using GT strain cells (\circ , \bullet), Δdh mutant cells (\square , \blacksquare) and $\Delta adh\Delta dh$ mutant cells (\triangle , \blacktriangle) in HEPES buffer solutions (shown by open symbols) and HEPES buffer solutions with 50 $\mu\text{mol mL}^{-1}$ fructose (shown by solid symbols). (a) y_{H_2} , number of moles hydrogen per culture volume, (b) c_s , concentration of unreacted exogenous fructose, (c) m_G , number of moles of endogenous glucose per dry cell weight, (d) X , concentration of dry cell weight, (e) c_L , concentration of lactate and (f) c_A , concentration of acetate. Gas phase was nitrogen gas. All experiments were carried at least in duplicate

The curves of y_{H_2} versus time in Figure 2(a) show that fructose increased initial hydrogen production rate and stabilized and accelerated hydrogen production rate after the initial stage. The initial hydrogen production rates in GT strain, Δdh mutant and $\Delta adh\Delta dh$ mutant, which were 0.025, 0.015 and 0.020 $\mu\text{mol mL}^{-1} \text{h}^{-1}$, respectively, in the absence of fructose, were increased to 0.091, 0.109 and 0.129 $\mu\text{mol mL}^{-1} \text{h}^{-1}$, respectively, in the presence of fructose. Fructose is found to largely enhance NiFe-hydrogenase activity. The hydrogen production rates after initial stage in GT strain, Δdh mutant and $\Delta adh\Delta dh$ mutant were declined with time as shown by Eq.(3). The hydrogen production in GT strain in fructose shown in Figure 2(a) is related by

$$r_{H_2} = r_{H_2,0} \quad (4)$$

with the deactivation constant k at 0 h^{-1} . The rates of hydrogen production in Δaddh mutant in fructose from 0 h to 84 h and in $\Delta\text{adh}\Delta\text{addh}$ mutant in fructose from 0 h to 72 h are also related by Eq.(4). After that hydrogen production in Δaddh mutant and $\Delta\text{adh}\Delta\text{addh}$ mutant in fructose were accelerated. Details concerning fructose effect and dehydrogenase elimination effect on hydrogen production are not entirely clarified, however, presence of exogenous fructose or elimination of genes coding for NAD(P)H consuming dehydrogenases appears related to metabolic reactions that continuously generate and supply electrons and NAD(P)H to NiFe-hydrogenase. If t_c is the transition time from stable hydrogen production to accelerated hydrogen production, t_c values for Δaddh mutant and $\Delta\text{adh}\Delta\text{addh}$ mutant are 84 and 72 h, respectively. The y_{H_2} values at 120 h of runs with fructose for GT strain, Δaddh mutant and $\Delta\text{adh}\Delta\text{addh}$ mutant were 10.8, 16.2 and 18.7 $\mu\text{mol mL}^{-1}$, respectively. This numbers were very much improved from non-substrate fermentation conditions which yielded hydrogen gas as 1.66, 1.00 and 1.42 $\mu\text{mol mL}^{-1}$ from GT, Δaddh and $\Delta\text{adh}\Delta\text{addh}$ strains, respectively.

In Figure 2(b), the affinity of derivatives from GT strain to fructose is extremely low when compared to that to glucose (data not shown). Fructose was consumed by cells of GT strain, Δaddh mutant and $\Delta\text{adh}\Delta\text{addh}$ mutant for maintaining cell mass stability. The consumption of fructose by GT, Δaddh and $\Delta\text{adh}\Delta\text{addh}$ strains were comparable. Consumption of fructose during 120 h was 5.1-8.4 $\mu\text{mol mL}^{-1}$. High hydrogen production of those cells in the presence of fructose was not likely resulted from direct fructose consumption.

In Figure 2(c), the time courses of the amount per culture volume of glycogen in terms of endogenous glucose (m_G) was displayed. In the first 24 h of dark incubation in HEPES buffer solution without or with fructose, the m_G increased in GT strain, Δaddh mutant and $\Delta\text{adh}\Delta\text{addh}$ mutant. This amount of endogenous glucose is corresponding to consumption of fructose. The peak value of m_G of runs with fructose is higher than that without fructose. The $\Delta\text{adh}\Delta\text{addh}$ mutant converted more fructose to endogenous glucose than GT strain and Δaddh mutant. The glycogen content dropped during dark anaerobic fermentation. The highest drop in $m_G X$ from 24 h to 120 h was 0.183 $\mu\text{mol mL}^{-1}$ that was observed in run with $\Delta\text{adh}\Delta\text{addh}$ mutant in fructose solution. Hydrogen generated in this run is 25.5 times that estimated from the theoretical yield of NAD(P)H in glycolysis. Cells used fructose for gluconeogenesis in the first 24 h then for glycolysis from glycogen after 24 h. At the end of fermentation, all strains had the same level of glycogen content.

In Figure 2(d), the time courses of the dry cell weight concentration of cyanobacterium cells (X) was illustrated. The dry cell weight concentration of GT strain, Δaddh mutant and $\Delta\text{adh}\Delta\text{addh}$ mutant slowly decreased with time during 120 h in the absence of fructose. The aqueous solution for cell suspension was consisted of milliQ grade water and HEPES-KOH for pH buffering. Nitrate was totally removed from the system. The buffer solution is confirmed to be insufficient to support cell viability of GT strain, Δaddh mutant and $\Delta\text{adh}\Delta\text{addh}$ mutant. In the presence of fructose, the X of GT strain increased in the first 24 h, was constant between 48 h and 72 h and increased again after 72 h, the X of Δaddh mutant was constant in the first 48 h, increased between 48 h and 72 h, decreased between 72 h and 96 h and increased after 96h, and the X of $\Delta\text{adh}\Delta\text{addh}$ mutant was constant for the first 72 h and increased after 72 h. The dry cell weight concentrations of GT strain, Δaddh mutant and $\Delta\text{adh}\Delta\text{addh}$ mutant at 120 h were higher than the initial concentrations. Curves for runs with fructose in Figures 2 (c) and (d) show that GT strain, Δaddh mutant and $\Delta\text{adh}\Delta\text{addh}$ mutant are capable of limited heterotrophic growth on fructose in HEPES buffer solution. In the presence of fructose, the X at 120 h was highest in runs with GT strain and Δaddh mutant and lowest that with $\Delta\text{adh}\Delta\text{addh}$ mutant. It is observed that growth period of Δaddh mutant is later than that of GT strain, and that of $\Delta\text{adh}\Delta\text{addh}$ mutant is later than that of Δaddh mutant. Instability of dry cell weight concentration in dark anaerobic nitrate-free HEPES buffer solution was suppressed by fructose. This result shows that fructose supports cell viability in the dark anaerobic condition.

In Figure 2(e), the time courses of lactate concentration (c_L) are shown. The production of lactate by GT strain in the presence of fructose is extremely high. Lactate production of GT strain in the presence of fructose was activated at 72 h. At 120 h, lactate concentration of run with GT strain, which was 0.204 $\mu\text{mol mL}^{-1}$ in buffer solution, increased to 9.02 $\mu\text{mol mL}^{-1}$ in the presence of fructose. Photosynthetic growth for cell preparation and dark anaerobic incubation were performed in the absence of antibiotics, hence the observation of trace amount lactate in runs with Δaddh mutant and $\Delta\text{adh}\Delta\text{addh}$ mutant shows the drop of phenotype of lactate dehydrogenase deficiency.

In Figure 2 (f), the time courses of acetate concentration (c_A) were plotted. Acetate concentrations of runs with fructose were higher than those without fructose, and increased drastically during 48-72 h. In the presence of fructose, acetate concentration was highest in GT strain, second highest in Δaddh mutant and lowest in $\Delta\text{adh}\Delta\text{addh}$ mutant. Acetate concentrations were highest during 48-72 h then the concentration dropped. The peaks of acetate concentration in runs with buffer solutions were 0.062, 0.162, and 0 $\mu\text{mol mL}^{-1}$ for GT strain, Δaddh mutant and $\Delta\text{adh}\Delta\text{addh}$ mutant, respectively, and those with buffer solutions with fructose were 1.21, 0.847 and 0.394 μmol

mL^{-1} for them, respectively. The concentration of acetate after 72 h was extremely low. The consumption of acetate is highest in run with GT strain, second highest in run with Δddh mutant and lowest in run with $\Delta\text{adh}\Delta\text{ddh}$ mutant. This order is reverse of that of hydrogen production, while close to that of dry cell weight production. It suggests that acetate consumption after the peak seems related to late cell growth

Ethanol concentrations in runs without or with fructose were not in detachable level. Our previous report shows the ethanol production of GT strain, Δddh mutant and $\Delta\text{adh}\Delta\text{ddh}$ mutant are 0.70, 1.01 and 0.47 $\mu\text{mol mL}^{-1}$, respectively, in the presence of glucose after 96 h (Chongsuksantikul et al., 2014), hence fructose fermentation of above strains appears to be different from glucose fermentation.

High production of hydrogen, endogenous glucose and acetate were observed if fructose was present. This observation raised very interesting points concerning participation of fructose in NAD(P)H production or redirection of NAD(P)H in the dark. The slight higher fructose consumption by mutants could be the reason that mutants took long time for cell growth therefore cell kept utilizing more carbon source. However at 120 h, difference of final dry cell weight concentrations among three strains was not large. Mutants consumed more fructose while produced less acid than GT strain. The m_G of Figure 2(c) is relatively stable in this period. More trials are required to find the products from acetate.

Cellular state variables of GT strain, Δddh mutant and $\Delta\text{adh}\Delta\text{ddh}$ mutant are summarized in Table 2, The initial hydrogen production rates of Δddh mutant and $\Delta\text{adh}\Delta\text{ddh}$ mutant increased 3-fold to 7-fold over GT strain when fructose was present. Hydrogen production from mutant lacking both *ddh* and *adh* genes yielded extremely high amount of hydrogen.

Table 2. Fructose-mediated dark anaerobic hydrogen production of Δddh mutant and $\Delta\text{adh}\Delta\text{ddh}$ mutant of *Synechocystis* sp. strain PCC6803. That of GT strain is also shown as a reference. Δddh , GT strain derivative lacking lactate dehydrogenase; $\Delta\text{adh}\Delta\text{ddh}$ mutant, GT strain derivative lacking both lactate dehydrogenase and alcohol dehydrogenase. Data of runs without or with 50 $\mu\text{mol mL}^{-1}$ fructose were analyzed. The $y_{\text{H}_2,1}$ was a number of moles of hydrogen per unit volume of buffer solution after 120 h

Strain	Growth phase of inoculum	Solution	X_0 [mg mL^{-1}]	$r_{\text{H}_2,0}$ [$\mu\text{mol mL}^{-1}\text{h}^{-1}$]	$y_{\text{H}_2,1}$ [$\mu\text{mol mL}^{-1}$]	K [h^{-1}]
GT strain	Logarithmic		2.1	0.0252	1.66	0.0151
Δddh mutant	Logarithmic	HEPES buffer solution	2.2	0.0150	1.00	0.0115
$\Delta\text{adh}\Delta\text{ddh}$ mutant	Logarithmic		2.0	0.0202	1.42	0.0119
GT strain	Logarithmic	HEPES buffer solution with 50 $\mu\text{mol mL}^{-1}$ fructose	1.9	0.0911	10.8	0
Δddh mutant	Logarithmic		2.2	0.109	16.2	0
$\Delta\text{adh}\Delta\text{ddh}$ mutant	Logarithmic		2.0	0.129	18.7	0

The amount per culture volume of hydrogen produced from GT strain was extremely high when logarithmic growth phase cells were incubated in dark anaerobic with supplement of fructose. This result implied that cells prepared from logarithmic growth phase have higher ability to utilize fructose.

Previously we reported that the deletion of the genes coding for competitive enzymes to hydrogenase was effective for increasing the initial hydrogen production rate of cells prepared from late-logarithmic growth phase. Present study shows that, if fructose is present, mutant cells prepared not from late-logarithmic growth phase but from logarithmic growth phase produce highest amount of hydrogen. The selection of growth phase affects hydrogen production in mutants greatly. The presence of fructose significantly activates hydrogen production from mutants.

The overview of the data shown in Figure 2 indicates that genetic perturbation of redox status that is brought by disruption of genes coding for lactate dehydrogenase and alcohol dehydrogenase plays an integral part to increase hydrogen production. Deletion of lactate dehydrogenase activity and alcohol dehydrogenase activity results not only redirection of NAD(P)H molecule for lactate dehydrogenase and alcohol dehydrogenase to NiFe-hydrogenase but also reforming the redox homeostasis networks. The increased amount of hydrogen is

higher than that expected by above redirection of NAD(P)H. Genetically modified mutants appear to represent a cellular state under the added load of mutation. Fructose plays the other integral part to increase hydrogen production. Dry cell weight concentration of mutants is supported by fructose as that of GT strain. In the presence of fructose, cells of GT strain, Δ dh mutant and Δ adh Δ dh mutant express the redox balance systems to constantly make-up NAD(P)H molecules utilized by NiFe-hydrogenase. The supernumerary NAD(P)H synthesis in mutants appear to trigger the second reformation of the redox balance systems during 48-76 h. This reformation contributes to further increase the amount of hydrogen per culture volume. The second reformation occurs when acetate production is changed to acetate assimilation.

Our work has showed that fructose is important for hydrogen production in GT strain, Δ dh mutant and Δ adh Δ dh mutant. The low consumption of fructose suggests that the fructose-mediated hydrogen production is economical regarding fructose supply since the unreacted fructose after a batch hydrogen production can be utilized in a subsequent repeated-batch hydrogen production. Fructose is abundant in nature, hence the application of fructose to dark anaerobic hydrogen production of GT strain, Δ dh mutant and Δ adh Δ dh mutant in nitrate-free solution seems to be a good possibility.

Our observation that fructose supports cell concentration in the dark anaerobic nitrate-free condition is supported by a previous work that shows that fructose is a photoheterotrophic growth substrate of *Anabaena* (Ungerer et al. 2008). There are also previous reports that seem to contradict our findings. For phototrophic growth of *Synechocystis* sp. strain PCC 6803, fructose was reported to represent a toxicity (Flores & Schmetterer, 1986). Photoheterotrophic growth of *Synechocystis* sp. strain PCC 6714 and some of other cyanobacteria on fructose have been reported to be hardly observed (Rippka et al., 1979; Joset et al., 1988). Cyanobacterium is a photosynthetic organism utilizing carbon dioxide for a main carbon source, there are remarkably few data available on the dark anaerobic heterotrophic growth. This work has opened the dark anaerobic heterotrophic growth of *Synechocystis* sp. strain PCC6803.

4. Conclusion

The hydrogen production of the glucose tolerant mutant of *Synechocystis* sp. strain PCC6803 in dark anaerobic HEPES buffer solution is found to be largely elevated by supply of fructose and elimination of dehydrogenase genes. Dark anaerobic 120 h incubation of a mutant lacking both lactate dehydrogenase genes and alcohol dehydrogenase genes in HEPES buffer solution with fructose is found to increase the amount of hydrogen per culture volume 11-fold over that without fructose. Fructose is observed to shift-up the amount of endogenous glucose per dry cell weight and to increase the stability of dry cell weight. The mechanism by which perturbation in redox status is related to under fructose remains to be elucidated. This work has proven that deletion of dehydrogenase enzyme results in the redirection of more electrons to flow to hydrogenase and the supplement of fructose contributes to long-term supply of electrons. Genetic perturbation in redox status and fructose addition in HEPES buffer solution, which we first elucidated in this study, appear to be a very promising biological approach to elevate dark anaerobic hydrogen production in cyanobacteria. This result has yielded very high number of moles of hydrogen per culture volume as no less than $18.7 \mu\text{mol mL}^{-1}$ when 2 mg mL^{-1} dry cell weight is utilized. This can be considered for future commercialization of biological process for hydrogen production by cyanobacteria.

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