Combining Amino Acid and Vitamin D Supplementation with Exercise Training Increases Skeletal Muscle Mass and Prevent Bone Mineral Density Loss in Participants with Low Muscle Mass

Ha Cao Thi Thu¹,², Satoshi Kurose¹, Yaeko Fukushima¹, Nana Takao¹,³, Natsuko Nakamura³, Akiko Kitamura³, Kyoko Higurashi¹, Tomohiro Kamo¹, Sawako Yoshiiuchi³, Katsuko Onishi¹, Hiromi Tsutsumi¹ & Yutaka Kimura¹,³

¹ Department of Health Science, Graduate School of Medicine, Kansai Medical University, Japan
² Department of Adult Nutrition Counseling, National Institute of Nutrition, Hanoi, Vietnam
³ Health Science Center, Kansai Medical University Hirakata Hospital, Japan

Correspondence: Ha Cao Thi Thu, Department of Health Science, Graduate School of Medicine, Kansai Medical University, 2-5-1, Shinmachi, Hirakata, Osaka, 573-1191, Japan. Tel: 81-72-804-2334; Fax: 81-72-804-2821. E-mail: caoha@hirakata.kmu.ac.jp

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Abstract

This study evaluated the impact of exercise training with amino acid and vitamin D supplementation on muscle and bone mass in participants with low muscle volume. Twenty-nine Japanese participants (56-84 years old) were enrolled and assigned into the supplement (n=15) and non-supplement (n=14) groups. All participants underwent a 6-month exercise program. Supplements and nutrition support were provided to the participants in the supplement group for 12 weeks. Body composition and whole bone mineral density (BMD) were measured using dual energy x-ray absorptiometry. The outcomes, including body composition, whole BMD, and skeletal muscle mass index (SMI), were evaluated twice: pre- and post-intervention. The SMI was 6.51(6.28; 7.14) and 5.58 (5.24; 6.05) (kg/m²) in men and women, respectively. The average SMI change was 0.13% (-0.05%; 0.31%) and 2.33% (-0.88%; 5.48%); [mean (lower; upper quartile)]. The average BMD loss in the non-supplement group was -2.78%, and the BMD increased in the supplement group by 4.34%; there was an absolute difference between the two groups (p<0.05). After the intervention, serum myostatin was changed (p=0.001, non-supplement>supplement), serum vitamin D was increased (p=0.03; supplement>non-supplement), and BMD was maintained (p=0.03, supplement>non-supplement). There was a significant difference in the serum myostatin level at baseline and at 6-month in the non-supplement group, with a mean difference of 0.48 ng/ml (p=0.01). There was no significant improvement in the total lean mass, and handgrip strength. Resistance exercise combined with an amino acid supplement affects muscle and bone mass in the short-term intervention.

Keywords: amino acid supplement; body composition; bone mineral density; skeletal muscle index; myostatin; vitamin D

1. Introduction

Sarcopenia is characterized by a loss of muscle mass and strength related to aging, and leads to physical disability, the decline in muscle function, falling, fractures, and increased risk of mortality (Cruz-Jentoft et al., 2010). More importantly, sarcopenia is associated with diseases that increase over time, especially in aging people. Older adults experience a progressive, generalized loss of skeletal muscle and decrease in physical function, with an inherent risk of disability, poor quality of life, and death (Morley, Baumgartner, Roubenoff, Mayer, & Nair, 2001). Sarcopenia has become common worldwide and is associated with high financial costs; hence, it is important to treat sarcopenia to maintain muscle mass, improve strength, reduce the risk of falling, and increase the life span of the elderly population.

Many factors such as insufficient physical activities, chronic disease, and decreased muscle protein synthesis may lead to the loss of muscle mass (Iannuzzi-Sucich, Prestwook, & Kenny, 2002), but the potentially preventable strategy is based on nutritional deficiency and skeletal muscle dysfunction with targeted interventions (Fiatarone et al., 1994). Growing evidence suggests that insufficient protein, energy, and certain micronutrients...
intake may consequently lead to secondary sarcopenia (Cerri et al., 2015). Nutrition intake can positively impact protein synthesis; therefore, interventions for sarcopenia usually include nutrition. Increasing the recommended dietary intake (quantity) and the essential amino acids (quality) of dietary protein can stimulate muscle protein anabolism in elderly people (Houston et al., 2008; Børsheim et al., 2008).

Stimulating gene expression, enhancing muscle protein synthesis and facilitating neuromuscular function were achieved by increasing vitamin D intake (Wicherts et al., 2007; Ceglia & Harris, 2013), and improving muscle strength and balance (Latham et al., 2003; Muir and Montero-Odasso, 2011). Vitamin D intake also declined the inflammation related to the slowdown of muscle strength in the elderly population (Schaap et al., 2006). The recommended nutritional intake includes high-quality protein and vitamin D, which are necessary to minimize the loss of muscle mass and its adverse effects in elderly (Waters et al., 2010; Candow et al., 2012). The decline of muscle mass and muscle function was addressed by physical training (Latham et al., 2004). Adding dynamic physical activity to resistance exercise also contributes substantially to improve physical function (Liu and Latham, 2009). Age and a decrease in nutrition are related (Forbes et al., 2012), and nutrition is an important modulator of health and well-being in the elderly. Inadequate nutritional intake may play a significant role in the metabolic control system and the progression of many lifestyle-related diseases.

Therefore, we designed a study that combined amino acids and a vitamin D supplement with regular physical activity. We tested whether, compared with non-supplementation, supplementation would increase the skeletal muscle index (SMI) and muscle mass while improving strength, nutritional status, and physical function.

2. Methods

2.1 Ethical Considerations

The study procedure and protocol were approved by the Institutional Review Board of Kansai Medical University (ethical documentation number: 000013445), and the study was performed according to the ethical principles of the Declaration of Helsinki. All participants gave written informed consent to participate.

2.2 Study Design and Participants

The present study was designed as an interventional investigation with supplementation and control (non-supplementation) groups. We recruited 19 participants, with low muscle mass who visited our health science center. The inclusion criteria were: participants aged 56 years and older who completed our intervention program and those with an appendicular lean mass divided by height squared (SMI), of <7.77 and <6.12 kg/m² for men and women, respectively. We excluded the participants with severe liver, heart, and kidney dysfunction; thyroid disorders; cancers; arthritis; laboratory abnormalities; BMI > 25 kg/m², and who were treated with steroids or heparin (3 persons). In the later period, we also recruited 18 other participants according to the previous inclusion and exclusion criteria. Three patients dropped out of the study and were excluded. Ultimately, the study included 29 participants. The details of study design are shown in Figure 1.

![Figure 1. Flow diagram of study design](image-url)
2.3 Intervention Procedures

Participants were examined by a physician for a medical consultation, underwent nutritional counseling, and received instructions on how to perform the physical exercise. Direct interviews were conducted to review the participant’s lifestyle regarding the intervention, to educate them about healthy eating behavior, and to develop a personalized plan for nutritional intake to maximize the participant’s protein intake behavior. Participants were instructed to keep their energy intake to the healthy weight × 25-35 kcal/day, and protein intake to healthy weight × 1.2 g/kg/day (Bauer et al., 2013). Recommended dietary fat and other nutrients intake was in accordance with the "Dietary Reference Intakes for Japanese (2015)". Healthy weight was calculated for each participant using the standard body mass index of 22 (kg/m²):

\[
\text{healthy weight} = 22 \times \text{height}^2
\]

Qualified dieticians conducted the monthly nutritional counseling. Exercise therapy was administered beginning at week 2. Each patient visited the health science center eight times a month for physical training and was followed up at 6-month.

2.4 Exercise Therapy

Physical activity, including a gravity level resistance exercise, was prescribed. The exercise duration was 60 min per day at the anaerobic threshold intensity level, 2 days per week, at our health science center under the supervision of qualified health fitness trainers. Prior to undertaking the exercise, each participant was evaluated using a symptom-limited cardiopulmonary exercise to determine the anaerobic threshold and peak oxygen uptake. In addition, the participants were instructed on how to perform the exercises at home, and were provided with weight instruments to help them perform these exercises. All the participants adhered to the prescribed intervention.

2.5 Outcome Measurement

The outcomes were evaluated according to the data collected from the interviews, muscle mass assessments, and physical function tests at baseline and at the 6-month follow-up interval. The participant’s body weight was measured on an electronic scale while he or she was barefoot and wearing light weight clothing. Standing height was measured without shoes, using a portable stadiometer with the mandibular plane parallel to the floor. The BMI was calculated as the whole-body weight divided by height squared. The SMI was calculated as the sum of the lean mass of both arms and legs divided by height squared. The handgrip dynamometer was used for evaluating handgrip strength (HGS). The maximum strength of the dominant hand of each participant was measured three times, and the average value was recorded. The usual and fastest walking speed was measured by determining the time that was required to walk a 10-m distance. The leg strength was evaluated using a recumbent ergometer (Strength Ergo; Mitsubishi Electric Corp., Tokyo, Japan). The maximum strength of the left and right legs of each participant was measured, and then the average value was calculated. Finally, these values were corrected for body weight (Nm/kg). During the test, recumbent ergometer settings were changed for each subject for standardization.

2.6 Laboratory Measures

Blood samples were obtained from the cubital vein via plastic tubes that were enclosed in a vacuum, between 8:30 and 10:00 AM after the participant had fasted for 8 h. We measured the peripheral and biochemical data, as well as myokines. The blood test was performed using a hospital autoanalyzer and it included: complete blood cell counts; neutrophil counts; hemoglobin, albumin, and fasting plasma glucose concentrations; estimated GFR, and lipid profile. The blood samples were centrifuged at 4 °C and 3,000 rpm for 10 min and placed in a container for serum storage at −80 °C until an enzyme-linked immunosorbent assay for serum myostatin (GDF-8/Myostatin Quantikine ELISA Kit, DGDF80, R&D Systems, Minneapolis, MN, USA) was performed using a Powerscan HT plate reader (DS Pharma Biomedical Co., Ltd., Osaka, Japan) and a chemiluminescent immunoassay for vitamin D (LIAISON 25 OH Vitamin D Total, DiaSorin S.p.A, Saluggia, Italy).

2.7 Body Composition

The whole-body composition was measured using dual energy x-ray absorptiometry (DXA) (QDR 4,500A, Hologic Inc., Bedford, MA, USA). The coefficient of variation for the lean mass measurements and bone mineral density was less than 3% and 1.5%, respectively. The fat and lean mass data and whole-body bone data were derived from a whole-body scan. The bone mineral content (BMC), fat mass, and lean soft tissue mass were measured separately for each part of the body, including the arms and legs. The SMI was calculated as the total mass of the arms and legs in the kilogram, divided by height in meters squared. All measurements were performed by the same technicians who used a standard operating procedure. The machine was calibrated using a standard
phantom before each measurement.

2.8 Hand Grip Strength

HGS was obtained using a handgrip dynamometer (Sammons Preston Rolyan Co, Bolingbrook, IL, USA). The HGS of each participant was measured three times for the maximum strength of the right and left hands, and then the average value was calculated.

2.9 Dietary Supplement

Amino acid supplements were provided to the participants in the supplement group every 2 weeks. Packets of jelly amino acid supplements, “Amino L40” (Ajinomoto, Tokyo, Japan), with 3 g of amino acids (40.0% leucine, 10.0% isoleucine, 10.0% valine, and 40% other) and 800 IU of vitamin D were provided to the participants, who were instructed to take the 3-g supplement twice daily (6 g) for 12 weeks after breakfast and dinner. To monitor the participant’s amino acid intake accurately, we provided them with record sheets that were collected every 2 weeks. Each participant recorded the amount and the time the supplement was taken every day.

2.10 Statistical Analysis

The statistical analysis of the data was designed to address the study's primary aims. The characteristics of the participants and other variables are presented as medians (lower and upper quartiles). The non-normal distribution data were checked by using the Shapiro-Wilk test. Nonparametric tests were performed using the Wilcoxon-signed rank test to determine the difference between the baseline and 6-month follow-up data, and the Wilcoxon rank sum test was used to examine the differences between the two groups. The proportion of participants who smoke, drink, and use drug was presented using the chi-square test, which determined the difference between the two groups. Spearman’s rank correlation coefficients were used to explore associations between myostatin and other variables. Percent change of variables was calculated using the formula:

\[
\text{% change} = \left( \frac{\text{[6-month value} - \text{baseline value]}}{\text{[baseline value]}} \times 100 \right)
\]

P-values less than 0.05 were considered statistically significant. All analyses were performed using the R statistical package (R Development Core Team 2008).

3. Results

The patients’ characteristics at baseline were similar between the groups for all factors. The main characteristics of the participants are shown in Table 1. The changes in variables between baseline and 6-month in the two groups are shown in Table 2. After the 6-month intervention program, there was a significant increase in the SMI in the supplement group (p=0.02), but the SMI in the non-supplement group did not exhibit a significant change. The mean serum myostatin level (483.78 ng/ml) increased significantly in the non-supplement group (p=0.01), but it did not significantly increase in the supplement group (p=0.1). The change of the serum myostatin level was significantly different between the two groups (p=0.001). There was no significant relationship between the change in myostatin level and the change in SMI scores, bone mineral density, and total lean mass. The serum vitamin D level increased significantly (p=0.03; supplement > non-supplement). The BMD and BMC did not change significantly from baseline to the 6-month follow-up in both groups, but there was a significant difference of BMD between the two groups (p=0.03, supplement > non-supplement); Figure 2. The SMI response was significantly different between baseline and at the 6-month interval, with a difference of 2.52% (0.64%; 5.24%) [median (lower; upper quartile)] in the supplement group (p=0.02), but not in the non-supplement group (p=0.43). Leg strength also showed the crucial change between baseline and at the 6-month interval in the supplement group (p=0.05), and in the non-supplement group (p=0.04). We did not see any significant improvement in the total lean mass and HGS in either group. No participants experienced an adverse event after taking the amino acid and vitamin D supplements. All participants complied with the intervention.
Table 1. Comparison of the baseline characteristics of selected variables between the non-supplement and supplement participants

<table>
<thead>
<tr>
<th>Variable</th>
<th>Non-supplement</th>
<th>Supplement</th>
<th>P-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>n</td>
<td>14</td>
<td>15</td>
<td></td>
</tr>
<tr>
<td>Age (years)</td>
<td>68.00 (60.50-72.75)</td>
<td>74.00 (67.00-77.00)</td>
<td>0.20</td>
</tr>
<tr>
<td>Height (m)</td>
<td>1.56 (1.49-1.62)</td>
<td>1.59 (1.53-1.61)</td>
<td>0.45</td>
</tr>
<tr>
<td>Body weight (kg)</td>
<td>58.80 (46.78-70.00)</td>
<td>53.40 (46.50-58.80)</td>
<td>0.45</td>
</tr>
<tr>
<td>BMI (kg/m²)</td>
<td>23.78 (19.36-26.15)</td>
<td>22.08 (18.71-23.58)</td>
<td>0.20</td>
</tr>
<tr>
<td>SBP (mmHg)</td>
<td>136.50 (125.75-142.00)</td>
<td>130.00 (115.00-169.00)</td>
<td>0.88</td>
</tr>
<tr>
<td>DBP (mmHg)</td>
<td>75.00 (72.00-85.25)</td>
<td>74.00 (70.00-95.00)</td>
<td>0.81</td>
</tr>
<tr>
<td>Total lean mass (kg)</td>
<td>34.95 (30.54-43.89)</td>
<td>34.75 (31.82-41.19)</td>
<td>0.85</td>
</tr>
<tr>
<td>Legs muscle mass (kg)</td>
<td>11.05 (9.56-13.75)</td>
<td>11.79 (9.97-13.07)</td>
<td>0.75</td>
</tr>
<tr>
<td>Arms muscle mass (kg)</td>
<td>3.13 (2.85-4.16)</td>
<td>3.39 (2.87-4.14)</td>
<td>0.71</td>
</tr>
<tr>
<td>Skeletal muscle index</td>
<td>6.50 (5.40-6.87)</td>
<td>6.15 (5.17-6.51)</td>
<td>0.31</td>
</tr>
<tr>
<td>Hand grip strength (kg)</td>
<td>21.00 (18.31-24.55)</td>
<td>20.50 (17.70-26.83)</td>
<td>0.91</td>
</tr>
<tr>
<td>Leg strength (N/kg)</td>
<td>1.25 (1.11; 1.29)</td>
<td>1.26 (1.09; 1.59)</td>
<td>0.75</td>
</tr>
<tr>
<td>Usual walking speed (m/s)</td>
<td>1.33 (1.30-1.39)</td>
<td>1.38 (1.20-1.58)</td>
<td>0.29</td>
</tr>
</tbody>
</table>

**Smoking status**

<table>
<thead>
<tr>
<th></th>
<th>Non-supplement</th>
<th>Supplement</th>
<th>P-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Never</td>
<td>12(41.4%)</td>
<td>14(48.3%)</td>
<td>0.571</td>
</tr>
<tr>
<td>Current</td>
<td>1(3.4%)</td>
<td>0</td>
<td></td>
</tr>
<tr>
<td>Past</td>
<td>1(3.4%)</td>
<td>1(3.4%)</td>
<td></td>
</tr>
</tbody>
</table>

**Drinking status**

<table>
<thead>
<tr>
<th></th>
<th>Non-supplement</th>
<th>Supplement</th>
<th>P-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Never</td>
<td>12(41.4%)</td>
<td>14(48.3%)</td>
<td>0.501</td>
</tr>
<tr>
<td>Current</td>
<td>2(6.9%)</td>
<td>1(3.4%)</td>
<td></td>
</tr>
<tr>
<td>Past</td>
<td>0</td>
<td>0</td>
<td></td>
</tr>
</tbody>
</table>

**Drug usage**

<table>
<thead>
<tr>
<th></th>
<th>Non-supplement</th>
<th>Supplement</th>
<th>P-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Hypertension</td>
<td>2(6.9%)</td>
<td>4(13.8%)</td>
<td>0.411</td>
</tr>
<tr>
<td>Hyperlipidemia</td>
<td>1(3.4%)</td>
<td>1(3.4%)</td>
<td>0.960</td>
</tr>
</tbody>
</table>

Notes. BMI, Body mass index; SBP, Systolic blood pressure; DBP, Diastolic blood pressure; Values shown are median, lower, and upper quartiles, and percent (in parentheses). P-values were derived from Wilcoxon rank sum test and chi-square test for difference between the two groups.
Table 2. Comparison of selected variables between the non-supplement and supplement groups at baseline and at 6-month follow-up

<table>
<thead>
<tr>
<th>Variable</th>
<th>Non-supplement</th>
<th>Supplement</th>
<th>6th month</th>
<th>6th month</th>
</tr>
</thead>
<tbody>
<tr>
<td>n</td>
<td>14</td>
<td>15</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Body weight (kg)</td>
<td>55.80 (46.78; 70.0)</td>
<td>56.25 (46.95; 66.10)</td>
<td>53.40 (46.50; 58.80)</td>
<td>54.00 (46.50; 56.70)</td>
</tr>
<tr>
<td>SBP (mmHg)</td>
<td>136.50 (125.75; 142)</td>
<td>138.00 (123; 147.50)</td>
<td>130.0 (115.0; 169.0)</td>
<td>132 (124.75; 144)</td>
</tr>
<tr>
<td>DBP (mmHg)</td>
<td>75.00 (72.00; 85.25)</td>
<td>75.50 (71.75; 83.00)</td>
<td>74.00 (70.00; 95.00)</td>
<td>81.5 (67.50; 86.25)</td>
</tr>
<tr>
<td>BMI (kg/m²)</td>
<td>23.78 (19.36; 26.15)</td>
<td>23.27 (19.75; 25.94)</td>
<td>22.08 (18.71; 23.58)</td>
<td>21.53 (19.05; 23.83)</td>
</tr>
<tr>
<td>Leg strength (N/kg)</td>
<td>1.25 (1.11; 1.29)</td>
<td>1.39 (1.29; 1.53)*</td>
<td>1.26 (1.09; 1.59)</td>
<td>1.48 (1.16; 1.66)*</td>
</tr>
<tr>
<td>Total muscle mass (kg)</td>
<td>34.95 (30.54; 43.89)</td>
<td>36.64 (31.43; 42.17)</td>
<td>34.74 (31.81; 41.19)</td>
<td>36.15 (32.56; 40.91)</td>
</tr>
<tr>
<td>Arms muscle mass (kg)</td>
<td>3.03 (2.88; 3.79)</td>
<td>3.29 (2.86; 4.09)</td>
<td>3.19 (2.87; 4.14)</td>
<td>3.47 (2.94; 4.17)*</td>
</tr>
<tr>
<td>Legs muscle mass (kg)</td>
<td>10.45 (9.41; 12.67)</td>
<td>10.58 (9.37; 13.11)</td>
<td>11.35 (9.97; 12.87)</td>
<td>11.60 (10.36; 13.21)*</td>
</tr>
<tr>
<td>SMI (kg/m²)</td>
<td>5.77 (5.41; 6.21)</td>
<td>6.08 (5.38; 6.47)</td>
<td>6.15 (5.18; 6.51)</td>
<td>6.40 (5.38; 6.70)*</td>
</tr>
<tr>
<td>BMC (g)</td>
<td>1860 (1647.50; 2381.25)</td>
<td>1787 (1456.25; 2327.75)</td>
<td>1893 (1550; 2090)</td>
<td>1977 (1623; 2600)</td>
</tr>
<tr>
<td>Vitamin D (ng/ml)</td>
<td>14 (13; 18.75)</td>
<td>16 (13.75; 21)</td>
<td>17 (14; 20)</td>
<td>26 (18; 31)**</td>
</tr>
<tr>
<td>Myostatin (ng/ml)</td>
<td>2.83 (2.09; 3.06)</td>
<td>3.26 (2.76; 3.56)*</td>
<td>2.60 (2.31; 3.59)</td>
<td>2.41 (2.14; 2.83)</td>
</tr>
</tbody>
</table>

Notes. SBP, Systolic blood pressure; DBP, Diastolic blood pressure; BMI, Body mass index; SMI, Skeletal muscle index; BMC, Bone mineral content; Values shown are median, lower and upper quartiles (in parentheses). *P<0.05, **P<0.01, P-values were derived from Wilcoxon signed-rank test for difference between baseline and 6-month.

Figure 2. Mean changes in vitamin D, myostatin, skeletal muscle index, total lean mass, bone mineral density, bone mineral content after the intervention. Bars indicate average changes from baseline to the 6-month follow-up; error bars represent the lower and upper quartile. BMD, bone mineral density; SMI, skeletal muscle index; LM, total lean mass; LML, lean mass legs. *P<0.05, **P<0.01, P-values were derived from Wilcoxon rank sum test for difference between the two groups.
4. Discussion

In this study, low muscle volume was defined based on the declines in the SMI. Our discussion focuses on the effects of the intervention as exploring pre-sarcopenia was the initial aim of this investigation. Many researchers performed investigations that focused on exercise and nutritional intake to control the decline of muscle mass and strength in patients with sarcopenia (Campbell, Crim, Dallal, Young, & Evans, 1994; Arnal et al., 1999; Kim et al., 2012; Rondanelli et al., 2016), but the results of these studies have not always been consistent (Kerstetter et al., 2005; Dillon et al., 2009; Paddon-Jones & Rasmussen, 2009). Our study demonstrates that the arm and leg muscle mass, hand grip strength, and walking speed did not increase with combined exercise and amino acid ingestion in both groups; however, the SMI improved only in the exercise and amino acid supplement group.

The Recommended Dietary Allowances (RDA) for dietary protein is based on the level of protein that is required to achieve nitrogen equilibrium; it was originally 1.0 g/kg until the 1980 RDA guideline and was subsequently decreased to 0.80 g/kg. Significant evidence suggests that the current recommendations for dietary protein in elderly people (≥65 years old) are inadequate for optimal muscle and bone health (Campbell et al., 1994). These studies assessed the effects of different amino acid doses (between 6.70 and 20 g/day) on protein synthesis, and the 6.0-g/day dose that was provided in our study is lower than that in previous studies; however, the mean weight of the subjects in those studies was between 71.0 and 81.3 kg, and the amino acid dose was between 0.09 and 0.25 g/kg of the participants’ weight. Our study used an amino acid dose of 0.11 g/kg, which is comparable with the doses used in the literature. Our study showed that the SMI was significantly increased in the supplement group, which is consistent with results of the previous investigations.

The increase in muscle mass due to nutritional supplementation was indicated in many studies; however, an increase in muscle strength does not always follow an increase in muscle mass. Although the current research showed that amino acid supplements combined with exercise increased the SMI and serum vitamin D level, as well as reduced the level of myostatin, an enhancement in muscle strength was observed in the leg strength. The improvement in single variables, such as leg muscle mass or the usual walking speed, knee extension strength can be observed in the exercise without supplement group (Peterson et al., 2010; Kim et al., 2012). However, rationally, improvements in single variables are not sufficient to treat sarcopenia. Although it is inconclusive whether exercise combined with an amino acid supplement is better than either intervention alone, these results suggest that exercise and amino acid supplementation may be necessary to gain muscle mass and muscle power.

This study has novel aspects due to its focus on body composition, bone mass, and supplement use. Elderly people must increase the anabolic stimulus and consume 30 g of protein per meal to prevent protein catabolism (Dillon et al., 2009; Paddon-Jones et al., 2009). The amino acid leucine stimulates skeletal muscle protein synthesis (Volpi et al., 1998). An increase in myostatin concentrations inhibits muscle growth. This is because myostatin is possibly the most predominant mediator of muscle growth and repair. Furthermore, in our study, the non-supplement participants exhibited a significant increase in the mean serum myostatin level (p=0.02), which may be due to the fact that the protein synthesis rate was low in this group. The myostatin level and the protein synthesis rate may be contributed to the building of muscle skeletal mass. Resistance training promoted significant enhancements in muscle strength, muscle power in elderly (Lopez et al., 2017), but not muscle skeletal mass. Otherwise, non-supplement participants may have low protein intake due to the absence of nutrition support and supplement. The expression of myostatin increment to restrict the enhancement of muscle tissue amid the procedure of muscular growth (Han et al., 2011). We did not find any significant relationship between myostatin and the SMI, which may be due to the fact that the dose of myostatin and the myostatin receptor were not well coordinated.

A recent meta-analysis (Beaudart et al., 2014) indicated the growth of muscle strength by 17% on average due to daily supplementation with 400 IU of vitamin D3. The supplement that we used contained 200 mcg (800 IU) of vitamin D. We used this dose of vitamin D because the mean age of the patients was 70, and the dose indicated for this age group is 800 IU of vitamin D daily to prevent bone loss; furthermore, these participants also participated in indoor training (Cranney et al., 2008).

The present study’s findings must be interpreted within the context of its strengths and potential weaknesses. The participant’s body composition was measured using DXA technology, which is considered the gold standard method for assessing body composition. We also measured the serum vitamin D and myostatin levels, which could allow for a better evaluation of muscle and bone metabolism. However, we used a small sample size, and we could not conduct subgroup analyses to examine the effect of the intervention in smaller groups of participants. Our study also did not have non-exercise and non-supplement groups to act as controls; otherwise, the effect of home exercise was not estimated, therefore, it is possible that the effect observed here may be an overestimation of the true effect sizes. Three people dropped out of the study, and they were excluded from the data analysis. However,
there were no significant differences between participants who withdrew from the study and those who completed the study. The duration of follow-up was only 6 months, and we were not able to ascertain the long-term effect of this intervention program. Furthermore, the effects of vitamin D supplement independently from amino acid supplement were not examined due to the small sample size.

5. Conclusion

The results of this study suggest that amino acid combined vitamin D supplement with resistance exercise can produce changes in catabolic and anabolic mediators, thereby lowering muscle inhibitor markers such as myostatin and improving serum vitamin D. This shift resulted in a significant increase in the SMI (+2.78%) and BMD (+4.34%), proving that the intervention is effective for treating low muscle mass and preventing sarcopenia, with improvements in the SMI and muscle power. Further follow-up studies that emphasize nutritional intake, with home exercise recording, with a bigger sample size, and having a control group without exercise are required to confirm these results.

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Authors' Contributions

HC responded for designing the study and wrote the manuscripts. HT, SY, AK, NT were responsible for monitoring the study and recording the data. SK, HC, KO were responsible for performing experiments and data analysis. KH, TK, YF, KN assisted in writing the manuscript. YK approved the final version.

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Competing Interests Statement

The authors declare that they have no conflicts of interest with respect to this paper.

References


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